- 1 Variability in trough total and unbound teicoplanin concentrations and achievement of
- 2 therapeutic drug monitoring targets in adult patients with haematological malignancy

3

- 4 Catherine J. Byrne¹, Jason A. Roberts^{2,3}, Brett McWhinney⁴, Jerome P. Fennell⁵, Philomena O'Byrne⁵,
- 5 Evelyn Deasy⁵, Sean Egan⁵, Ronan Desmond⁵, Helen Enright⁵, Sheila A. Ryder^{1#}, Deirdre M. D'Arcy^{1*},
- 6 Johnny McHugh^{5*}

7

- 8 ¹School of Pharmacy and Pharmaceutical Sciences, Trinity College Dublin, Dublin 2, Ireland; ²Burns,
- 9 Trauma and Critical Care Research Centre, The University of Queensland, Brisbane, Australia; ³Centre
- 10 for Translational Anti-Infective Pharmacodynamics, The University of Queensland, Brisbane,
- Australia; ⁴Queensland Pathology, Brisbane, Australia; ⁵Tallaght Hospital, Dublin 24, Ireland.

12

- 13 Running title: Teicoplanin therapy in haematological malignancy
- 14 **Keywords:** teicoplanin, haematological malignancy, therapeutic drug monitoring, pharmacokinetics,
- 15 protein binding

16

- [#]Corresponding author. School of Pharmacy and Pharmaceutical Sciences, Trinity College Dublin,
- Dublin 2, Ireland. Tel: +353 1 896 2786; Email: sryder@tcd.ie
- 19 *Equal contribution

<u>Abstract</u>

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

Objectives: To explore the following aspects of teicoplanin use in patients with haematological malignancy: early attainment of target trough concentrations with current high dose teicoplanin regimens; variability in unbound teicoplanin fractions; factors associated with observed total and unbound trough concentrations; efficacy and toxicity; and renal function estimation. Methods: This was a single-centre, prospective study. Trough samples were taken on Days 3, 4, 7 and 10. Total and unbound teicoplanin concentrations were determined using validated HPLC methods. Regression analyses were used to identify factors associated with trough concentration. Results: Thirty teicoplanin-treated adults with haematological malignancy were recruited. High interpatient variability in trough total and unbound concentrations was observed (coefficient of variation 43.8% and 66.1%, respectively, at 48 h). Despite higher than conventional dosages, the proportions of patients with a trough concentration ≥20 mg/L at 48 h and at 72 h were 16.7% and 37.9%, respectively. There was a significant negative association between renal function and trough concentrations attained at 48 h and at 72 h (P<0.05). In the absence of measured creatinine clearance, estimates using the Cockcroft-Gault (total body weight) equation could prove an acceptable surrogate. Unbound fractions of teicoplanin were highly variable (3.4-18.8%). Higher unbound fractions were observed in patients with low serum albumin concentrations. Teicoplanin was well tolerated. Conclusions: Higher teicoplanin loading doses than those in current use appear necessary. Increased dosing is needed in patients with increased renal function. High variability in protein binding supports the contention for therapeutic drug monitoring of unbound teicoplanin concentrations. EudraCT registration 2013-004535-72.

<u>Introduction</u>

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

Infection is one of the most common complications of chemotherapy-induced neutropaenia (1). Haematological malignancy patients have the greatest risk for severe neutropaenia, compared to solid tumour patients, because of the underlying disease as well as the severely myelosuppressive chemotherapy used for treatment (2). The increasing incidence of Gram-positive pathogens in these patients is well recognised and, as these pathogens are often meticillin-resistant, glycopeptide antibiotics, commonly teicoplanin or vancomycin, have an important role in their treatment (1). Teicoplanin is considered to be a useful alternative to vancomycin - it is equally effective, can be administered once daily and is associated with fewer side-effects (3). Indeed, surveys conducted in the UK and Ireland have found teicoplanin to commonly be the preferred choice for patients with haematological malignancy (4, 5). However, the emergence of teicoplanin-resistance is a significant concern (6-8) and, coupled with the impaired ability of neutropaenic patients to fight infection, makes it important to achieve adequate exposure rapidly (9). The ratio of the area under the concentration-time curve to the minimum inhibitory concentration (AUC/MIC) is thought to be the pharmacokinetic/pharmacodynamic index best correlating with glycopeptide efficacy (10-12). However, calculating AUC requires multiple samples and therefore trough concentrations are used as a surrogate marker to assess exposure in daily clinical practice (13). Whilst the Summary of Product Characteristics specifies a target trough concentration of ≥10 mg/L for most infections (14), a higher trough target has been advocated for haematological malignancy patients (15, 16). Indeed, the trough target recommended at Tallaght Hospital for teicoplanin in haematological malignancy patients is ≥20 mg/L, with higher than conventional doses specified to achieve this. As teicoplanin is highly protein bound (90-95%) (17), altered serum albumin concentrations may have variable effects on total and unbound concentrations (18). Knowledge of unbound concentrations may be more relevant than total concentrations to predict outcome as unbound concentrations are responsible for antimicrobial activity and correlate best with drug response (18).

Previous data have suggested that albumin concentrations play a major role in the variability of the unbound (free) fraction (FF) of teicoplanin (13, 19-21). Altered FFs of teicoplanin and a lack of correlation between unbound and total concentrations might also be expected in haematological malignancy patients where low albumin concentrations are common (22). We previously reported a mixed effects regression model explaining 52% of the variability in teicoplanin trough total concentrations in haematological malignancy patients and identified dose, day of therapy, renal function and a diagnosis of acute myeloid leukaemia (AML) as significant factors associated with trough total concentrations (16). However, due to the retrospective nature of that study, critical characteristics that might also affect trough concentrations were not available, including fluid balance, illness severity measures and measured creatinine clearance (CL_{CR}). Furthermore, there was a lack of consistency in both dosing and day of trough concentration measurements. The objectives of this study were: (i) to assess whether current high dosing regimens of teicoplanin result in attainment of the target trough concentration on Days 3 and 4; (ii) to determine the variability in FFs of teicoplanin; (iii) to identify factors associated with both total and unbound trough concentrations attained on Days 3 and 4; (iv) to describe efficacy and toxicity; and (v) to compare the performance of renal function estimation equations for estimating measured CL_{CR}.

87

88

90

91

92

93

94

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

Methods

89 **Setting**

This single-centre, prospective study was conducted at Tallaght Hospital, Dublin, Ireland. Ethical approval was obtained from the Tallaght Hospital/St James's Hospital Joint Research Ethics

Committee (REC reference 2013/12/01). The study protocol was authorised by the Health Products

Regulatory Authority (Clinical Trial Number CT 900/545/1) and the trial was registered with the

European Clinical Trials Database Registry (EudraCT number 2013-004535-72). The study was

95 conducted following the guidelines of the Declaration of Helsinki. Written informed consent was 96 obtained from all patients. 97 Study population 98 The inclusion criteria were: (i) diagnosed with a haematological malignancy; (ii) age ≥18 years; (iii) 99 treated with teicoplanin for >48 h; (iv) intravascular catheter present; and (v) written informed 100 consent obtained. The exclusion criteria were: (i) receiving renal replacement therapy; (ii) admitted 101 to the Intensive Care Unit; (iii) incapable of comprehending the nature and scope of the trial; and (iv) 102 blood sampling personnel/analyst/processing equipment not available. 103 Dosing regimen 104 Teicoplanin (Targocid®, Sanofi, Dublin, Ireland) was administered intravenously by slow bolus injection. 105 The hospital dosage regimen was 600 mg (or 800 mg if weight >80 kg) 12-h for three loading doses 106 followed by 600 mg (or 800 mg if weight >80 kg) once daily. However, prescribed dosing regimens were 107 at the discretion of treating physicians and the hospital dosage regimen was not always followed. 108 Blood sampling, handling, storage and measurement 109 Trough samples (24 h post-dose) were taken on Days 3 (48 h), 4 (72 h), and 7 and 10 (when 110 applicable). Samples were immediately refrigerated and centrifuged within 6 h at 3000 rpm for 111 10 min. The supernatant was stored at -80°C until analysis. Total and unbound teicoplanin 112 concentrations were determined using HPLC as described by Roberts et al (13). 113 Determination of CL_{CR} 114 Urine was collected over a 24 h period on Day 3. The volume of urine was measured and a 1 mL aliquot 115 stored at -80°C until analysis. Urine creatinine concentration was determined locally using an enzymatic 116 method performed on a Roche/Hitachi Cobas C702 AutoAnalyzer system (Roche Diagnostics GmbH, 117 Mannheim, Germany). Urine volume, serum creatinine concentration (Scr) on the day of the urine

collection and urine creatinine concentration were used to calculate the measured CL_{CR}.

MIC testing

118

The identification of isolates from study patients was determined locally by broth microdilution using a VITEK®2 system (bioMérieux UK Ltd., Basingstoke, UK) as per routine care. Teicoplanin MICs of Grampositive isolates from blood cultures taken from study patients were determined locally with MIC test strips (Liofilchem, Italy).

Additional clinical and demographic data including age, body weight, height, serum albumin

Additional data

concentration, blood counts, 24 h fluid balance on Day 3, and measures of illness severity including the Multinational Association for Supportive Care in Cancer (MASCC) risk-index score (23), and the Charlson co-morbidity index (24), were collected. If a laboratory value was missing on a particular day, the next closest value to that day was used, provided it was within 2 days of the missing value.

Ideal body weight (IBW) was estimated using the Devine equation (25). Body surface area (BSA) was estimated using the Mosteller equation (26). Estimated CL_{CR} (eCL_{CR}) was calculated using the Cockcroft-Gault equation with total body weight (TBW) (CG-TBW) and IBW (CG-IBW) (27), and the Jelliffe (JEL) equation (28). Estimated glomerular filtration rate (eGFR) was calculated using the four-variable modification of diet in renal disease equation, both unadjusted (MDRD) and adjusted for BSA (MDRDa) (29), and the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation (30).

Factors associated with trough concentrations attained

The relationship between patient factors and trough total concentrations at 48 h (trough_{48h-total}) and 72 h (trough_{72h-total}), and trough unbound concentrations at 48 h (trough_{48h-unbound}) and 72 h (trough_{72h-unbound}) attained, were assessed. Log trough concentrations were used for the dependent variable as the data were positively skewed. Independent variables tested included: age; haematological malignancy diagnosis; receipt of a bone marrow transplant; sickness severity scores; measured CL_{CR} (Day 3 only), eCL_{CR} and eGFR; serum albumin concentration; fluid balance and fluid input.

Step-wise incorporation of covariates was conducted for multivariate model development with cumulative dose (mg/kg) included in all models. Covariates that did not contribute to, or reduced the fit of, the model were removed sequentially and only significant covariates were retained. The target total

trough concentration was 20 mg/L, and the target unbound trough concentration was 1.5 mg/L, assuming 92.5% protein binding. These targets were based on those suggested from previously published studies (13, 15, 16). Comparison of renal function estimation equations The performances of renal function estimation equations for estimating measured CL_{CR} were compared. The CG-TBW, CG-IBW and MDRDa estimates were compared with measured CL_{CR} in mL/min. The MDRD, CKD-EPI and JEL estimates were compared with measured CL_{CR} in mL/min/1.73 m². Bias was assessed as the median difference, with positive values indicating overestimation of measured CL_{CR}. Precision was assessed as IQR for the differences. Accuracy was assessed as root mean square error and percent of estimates within 30% of measured CL_{CR} (30). Response to teicoplanin therapy Assessment of response to teicoplanin therapy was conducted using the same methods and definitions as previously described by Byrne et al (16). Nephrotoxicity and hepatotoxicity Nephrotoxicity was assessed by comparing Scr on the first and last days of teicoplanin therapy. Nephrotoxicity was defined as an increase in Scr of >0.5 mg/dL or ≥50% (31). Hepatotoxicity was assessed by comparing serum alanine transaminase (ALT) on the first and last days of teicoplanin therapy and was defined as an increase in ALT of >3 times the upper limit of normal or >3 times baseline if the level was abnormal on Day 1 (31). Statistical analyses All statistical analyses were conducted using IBM SPSS Statistics for Windows v. 22 (IBM Corp., Armonk, NY) or Minitab 16 Statistical Software (Minitab Ltd., Coventry, UK). Data were described as the mean ± SD or the median (IQR) for continuous variables, and as the number (%) for categorical variables. Either unpaired Student's t-test or non-parametric Mann–Whitney U-test was used to compare groups for continuous variables. Fisher's exact test was used to compare groups for categorical covariates. Correlation between continuous variables was evaluated using the Pearson correlation coefficient (r).

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

Simple and multiple regression analyses were used to assess the relationship between patient factors and trough concentrations. Statistical significance was defined as P<0.05.

Results

Thirty patients were recruited into the study. A summary of demographic and clinical characteristics of included patients are provided in Table 1. Coagulase-negative staphylococcal (CoNS) central line-associated blood stream infection (CLABSI) was the most common microbiologically documented infection occurring in the cohort (n=7, 33.3%). Three patients (10%) died during their admission and this was attributed to progression of the malignancy in all cases.

Dosing regimens

All 30 patients received three initial loading doses ranging from 330 mg to 800 mg (4.7-13.8 mg/kg).

Twenty nine patients received once daily maintenance doses of 600 mg or 800 mg

(7.3-13.8 mg/kg/day). One patient received 800 mg once daily (8.8 mg/kg) up to Day 8 and then twice daily thereafter. The duration of teicoplanin therapy ranged from 3-20 days.

Trough concentrations

High interpatient variability in trough total and unbound concentrations was observed. Fig. 1 summarises observed total and unbound trough concentrations on Days 3, 4, 7 and 10, and illustrates the accumulation of total and unbound teicoplanin over time.

The proportions of patients with a trough_{48h-total} and trough_{72h-total} of \geq 20 mg/L were 16.7% (5/30) and 37.9% (11/29), respectively. The proportions of patients with a trough_{48h-unbound} and trough_{72h-unbound} of \geq 1.5 mg/L were 26.7% (8/30) and 37.9% (11/29), respectively. There was a moderate correlation between total and unbound trough concentrations at 48 h and at 72 h (r=0.721, P<0.001; and r=0.692, P<0.001, respectively) (Fig. 2). The FFs of teicoplanin showed high interpatient variation, with FFs ranging from 3.4-18.8%. Higher FFs were observed in patients with low serum albumin concentrations (Fig. 3).

Factors associated with trough concentrations attained

198 All 30 patients were included in analyses of 48 h trough concentrations. Twenty nine patients were 199 included in analyses of 72 h trough concentrations, with one patient excluded due to lack of trough measurement at this time. A CL_{CR} of 1 mL/min was assumed for one patient based on the urine 200 201 output of ~10 mL on this day. 202 Trough_{48h-total} 203 The only factors significantly associated with the log trough_{48h-total} were eGFR using the MDRD $(R^2=14.0\%, P<0.05)$ and MDRDa $(R^2=14.6\%, P<0.05)$ equations, with a negative relationship, and the 204 MASCC score (R^2 =17.2%, P<0.05), with a positive relationship. 205 The best multiple regression model, including cumulative dose (mg/kg), was as follows: 206 Log trough_{48h-total}=1.0200 + 0.0110 cumulative dose -0.0019 MDRD (R^2 =24.4%, P<0.05, VIF=1.00). 207 208 Trough 72h-total 209 All renal function measures were significantly negatively associated with the log trough 72h-total, including measured CL_{CR} (R^2 =33.5%, P<0.001), eGFR using the MDRD (R^2 =25.5%, P<0.01) and MDRDa 210 $(R^2=33.0\%, P<0.005)$ equations, and eCL_{CR} using the CG-TBW $(R^2=24.3\%, P<0.01)$ and CG-IBW 211 $(R^2=19.0\%, P<0.05)$ equations. IBW showed a significant negative association $(R^2=17.6\%, P<0.05)$ and 212 the MASCC score showed a significant positive association (R^2 =17.9%, P<0.05) with the 213 214 log trough_{72h-total}. 215 The best multiple regression model, including cumulative dose (mg/kg), was as follows: 216 Log trough_{72h-total} = 1.1100 + 0.0025 cumulative dose -0.0021 CL_{CR} + 0.0134 MASCC (R^2 =50.1%, P<0.005, VIF=1.17). 217 According to this model, for a standard 70 kg patient with a CL_{CR} of 70 mL/min and a MASCC score of 218 219 16, the estimated loading regimen to achieve a trough_{72h-total} of 20 mg/L is 900 mg (13 mg/kg) 12-h for three doses and then a further dose 24 h later. 220 221 Trough_{48h-unbound}

IBW was the only factor significantly associated with the log trough_{48h-unbound}, with a negative

relationship (R^2 =20.3%, P<0.05). No multiple regression models were considered acceptable.

222

Trough 72h-unbound

Simple regression showed that all renal function measures were significantly negatively associated with the log trough_{72h-unbound}, including measured CL_{CR} (R^2 =41.9%, P<0.001), eGFR using the MDRD $(R^2=16.2\%, P<0.05)$ and MDRDa $(R^2=23.5\%, P<0.01)$ equations, and eCL_{CR} using the CG-TBW $(R^2=20.8\%, P<0.05)$ and CG-IBW $(R^2=22.9\%, P<0.01)$ equations. IBW, fluid input and albumin concentration were also significantly negatively associated with the log trough_{72h-unbound} (R^2 =31.0%, P < 0.005; $R^2 = 22.3\%$, P < 0.05; and $R^2 = 16.3\%$, P < 0.05, respectively). The best multiple regression model, including cumulative dose (mg/kg), was as follows: Log trough_{72h-unbound} = 0.1810 + 0.0046 cumulative dose -0.0033 CL_{CR} (R^2 =43.8%, P<0.005, VIF=1.16). According to this model, for a standard 70 kg patient with a CL_{CR} of 70 mL/min, the estimated loading regimen to achieve a trough_{72h-unbound} of 1.5 mg/L is 900 mg (13 mg/kg) 12-h for three doses and then a further dose 24 h later. Comparison of renal function estimation equations Fig. 4 and Table 2 show the performance of renal function estimation equations relative to

Fig. 4 and Table 2 show the performance of renal function estimation equations relative to measured CL_{CR}. The MDRDa and MDRD equations had the lowest bias. The CG-TBW and JEL equations had the highest precision. Accuracy, in terms of the highest percentage of estimates within 30% of measured CL_{CR}, was highest for the JEL and CG-TBW equations. Accuracy, in terms of the smallest root mean square error, was highest for CKD-EPI and CG-TBW equations. Overall, the CG-TBW, JEL and CKD-EPI equations performed comparatively well for estimating measured CL_{CR}. A comparison of model-predicted versus observed trough_{72h-total} in study patients, using measured CL_{CR} and using eCL_{CR} calculated by the CG-TBW equation as a substitute for measured CL_{CR}, is provided in Fig. 5.

Response to teicoplanin therapy

Of the 30 febrile episodes, seven cases were deemed evaluable for assessment of response to teicoplanin and all were meticillin-resistant CoNS CLABSIs. Of these, there were four successful outcomes and three failures. The median time to failure was 8 days (range 3-14 days). Causes of

failure were persistence of fever in two cases and persistence of both fever and pathogen in one case. Central lines were retained in all successful cases but not in the three failures.

There was no significant difference in clinical or demographic factors between successful and failed cases. The mean ± SD trough total and unbound concentrations, and trough total/MIC and trough unbound/MIC ratios, were higher in successful than in failed cases, although the differences were

Adverse events

not statistically significant (Table 3).

Overall, teicoplanin was well tolerated. Four patients developed skin rash (13.3%), but in all cases other medications with known potential to cause skin rash were used concurrently. A severe hypersensitivity reaction developed in one patient, within minutes after the IV infusion commenced, despite being treated with teicoplanin on a previous admission without consequence.

Nephrotoxicity was observed in five patients (16.7%). Of these, four were co-treated with other potentially nephrotoxic drugs and most often this was an aminoglycoside. In the remaining case, the onset of acute kidney injury ensued 3 days before teicoplanin was commenced. There was no significant difference between the median (IQR, range) highest trough total concentration in cases with evidence of nephrotoxicity [30.2 mg/L (15.6 mg/L, 13.9-37.5 mg/L), n=5] and cases with no evidence of nephrotoxicity [29.8 mg/L (14.3 mg/L, 16.7-74.9 mg/L), n=25] (P=1.000). There was no significant difference between the mean (SD, range) duration of therapy in cases with evidence of nephrotoxicity [8 days (6 days, 3-14 days), n=5] and cases with no evidence of nephrotoxicity [10 days (4 days, 3-20 days), n=25] (P=0.565). There was no evidence of hepatotoxicity in the study cohort.

Discussion

The findings of this prospective study provide further evidence that higher loading doses of teicoplanin are needed in patients with haematological malignancy. For an average haematological malignancy patient, the regression models developed suggest that sequential loading doses of at

least 12 mg/kg would be needed to achieve early adequate exposure. Loading doses of 12 mg/kg 12-h for 3-5 doses are currently recommended for bone and joint infections to achieve trough concentrations of ≥20 mg/L (14). Adopting these dosing recommendations for haematological malignancy patients may be an appropriate consideration. Consistent with studies in other patient groups (13, 19-21), FFs of teicoplanin were highly variable in study patients, with higher FFs observed in patients with low serum albumin concentrations. In recent years, the importance of therapeutic drug monitoring (TDM) of unbound teicoplanin concentrations has been highlighted for critically ill and chronically ill patients (13, 20, 21). Given the observed variability in protein binding and the lack of a strong correlation between trough total and unbound concentrations, TDM of unbound teicoplanin concentrations may prove useful in the future. The regression analyses showed that renal function is an important consideration for appropriate initial teicoplanin dosing, which is in keeping with the findings of recently published studies (16, 32, 33). Although the impact of renal function on trough concentrations was stronger at 72 h compared to at 48 h, the results suggest that in patients with enhanced renal function, achieving target trough concentrations may be difficult unless very high loading doses, such as >20 mg/kg, are used. Measured CL_{CR} had a stronger association with trough concentrations than estimated values calculated using renal function estimation equations. Should measured CL_{CR} data not be available, given the experience of use in clinical practice and the results of the comparison of renal function estimation equations, eCL_{CR} calculated using the CG-TBW equation could be proposed as a surrogate for measured CL_{CR} in this patient group. In our previous retrospective study, a diagnosis of AML showed a significant negative association with trough total concentrations (16), although this was not found to be the case in the current study. This may have been due to the smaller sample size and lower number of AML patients in the current study (n=7) compared to the retrospective study (n=20). We postulated that AML patients may have different underlying pathophysiology compared to patients with other types of

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

haematological malignancy, including higher fluid loads, inflammation and/or severity of illness. Indeed in the current study, fluid input and MASCC score were significantly associated with trough concentrations. The MASCC score is a composite score, used to identify the risk of complications in febrile neutropaenic cancer patients, with lower scores indicating a higher risk of complications (23). It is possible that lower MASCC scores reflect altered pathophysiology and/or the use of supportive treatments, such as aggressive fluid therapy, in sicker patients, resulting in enhanced disposition of teicoplanin. Demonstrating a relationship between teicoplanin concentrations and clinical outcome would have been useful to guide practice in this patient group. However, establishing the efficacy of an individual antibacterial agent is difficult in neutropaenic patients because antibacterial treatment is often prescribed empirically and these patients are frequently on several antibacterial agents concurrently. Such was the case in the current study, with only seven patients being evaluable for assessment of teicoplanin efficacy and therefore no further insight into the appropriate trough target for teicoplanin in haematological malignancy patients was gained. Nevertheless, the mean \pm SD trough_{48h-total} of 18.6 \pm 12.3 mg/L and trough_{72h-total} of 22.8 \pm 15.2 mg/L, observed in successful cases in the current study, were consistent with previous studies suggesting a target trough of 20 mg/L (15, 16). Of course, the benefits of using higher teicoplanin doses to produce higher trough concentrations must be balanced against the potential risk of increased toxicity. In the current study, with trough concentrations ranging from 4.1-70.5 mg/L between Days 3 and 10, teicoplanin was well tolerated. Apart from the severe hypersensitivity reaction in one patient, none of the adverse events observed could definitely be attributed to teicoplanin. Furthermore, no relationship between trough concentrations and incidence of adverse events was observed. This study had several limitations. Firstly, the study was conducted in a single centre and the sample size was small. Secondly, no conclusions could be made about the relationship between drug exposure and clinical outcomes because there were too few microbiologically documented Gram-

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

positive infections. Thirdly, the current study does not allow statements to be made about the relationship between trough levels and toxicity outside the range observed in this study. Fourthly, therapeutic targets for teicoplanin are not well defined and therefore the dosing estimations may be different should new targets be determined in the future.

In conclusion, to achieve target trough concentrations early in therapy, higher loading doses of teicoplanin than those in current use appear necessary in patients with haematological malignancy.

Renal function is an important consideration for appropriate initial dosing of teicoplanin. Serum albumin concentration has a significant effect on unbound teicoplanin concentrations. High variability in protein binding supports the contention for TDM of unbound concentrations.

Acknowledgements

The authors are grateful to Eddie McCullagh, Michael Kelly, Elaine O'Mullane, Li Wah Kyaw Tun, Sharon Curran-Rae and the nursing staff from Webb and Maguire Wards of Tallaght Hospital (Dublin, Ireland) for their valuable assistance with conducting this research. Special thanks are due to Prof. Ross Crosby (Director of Biomedical Statistics, Neuropsychiatric Research Institute, Fargo, ND) for statistical advice. Jason Roberts would like to recognise funding from the Australian National Health and Medical Research Council for a Centre of Research Excellence (APP1099452) and a Career Development Fellowship (APP1048652).

Funding

This research was supported by the Meath Foundation (grant reference SEJMH). The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

Transparency declarations

- 353 Jason Roberts has partaken in consultancies/advisory boards for Infectopharm, Astellas, MSD and
- 354 bioMerieux; in lectures with MSD and investigator initiated grants with MSD and Cardeas.
- 355 All other authors have none to declare.

References

- 1. **Kosmidis CI, Chandrasekar PH.** 2012. Management of gram-positive bacterial infections in patients with cancer. Leuk Lymphoma **53:**8-18.
- 359 2. **Ahn S, Lee YS.** 2012. Predictive factors for poor prognosis febrile neutropenia. Curr Opin Oncol **24:**376-380.
- 36. Menichetti F, Martino P, Bucaneve G, Gentile G, D'Antonio D, Liso V, Ricci P, Nosari AM,
 362 Buelli M, Carotenuto M, et al. 1994. Effects of teicoplanin and those of vancomycin in initial
 363 empirical antibiotic regimen for febrile, neutropenic patients with hematologic malignancies.
 364 Gimema Infection Program. Antimicrob Agents Chemother 38:2041-2046.
- Ziglam HM, Gelly KJ, Olver WJ. 2005. A survey of the antibiotic treatment of febrile
 neutropenia in haematology units in the United Kingdom. Clin Lab Haematol 27:374-378.
- Byrne CJ, Egan S, D'Arcy DM, O'Byrne P, Deasy E, Fennell JP, Enright H, McHugh J, Ryder
 SA. 2014. Teicoplanin usage in adult patients with haematological malignancy in the UK and
 Ireland: Is there scope for improvement? European Journal of Hospital Pharmacy: Science
 and Practice 21:301-305.
- 371 6. Pagano L, Tacconelli E, Tumbarello M, Laurenti L, Mele L, Spanu T, Cauda R, Fadda G, Leone
 372 G. 1997. Teicoplanin-resistant coagulase-negative staphylococcal bacteraemia in patients
 373 with haematological malignancies: a problem of increasing importance. J Antimicrob
 374 Chemother 40:738-740.
- Falcone M, Giannella M, Raponi G, Mancini C, Venditti M. 2006. Teicoplanin use and
 emergence of Staphylococcus haemolyticus: is there a link? Clin Microbiol Infect 12:96-97.
- Ahlstrand E, Svensson, K., Persson, L., Tidefelt, U., Soderquist, B. 2011. Glycopeptide
 resistance in coagulase-negative staphylococci isolated in blood cultures from patients with
 hematological malignancies during three decades. Eur J Clin Microbiol Infect Dis 30:1349 1354.
- Jarkowski A, 3rd, Forrest A, Sweeney RP, Tan W, Segal BH, Almyroudis N, Wang ES,
 Wetzler M. 2012. Characterization of vancomycin pharmacokinetics in the adult acute myeloid leukemia population. J Oncol Pharm Pract 18:91-96.
- 384 10. **Craig WA.** 2003. Basic pharmacodynamics of antibacterials with clinical applications to the use of beta-lactams, glycopeptides, and linezolid. Infect Dis Clin North Am **17**:479-501.
- 386 11. Drusano GL. 2004. Antimicrobial pharmacodynamics: critical interactions of 'bug and drug'.
 387 Nat Rev Microbiol 2:289-300.
- 388 12. Eisenberg MJ, Okrainec K, Lefkovits J, Goudreau E, Deligonul U, Mak KH, Duerr R, Tsang J,
 389 Huynh T, Sedlis S, Brown DL, Brieger D, Pilote L. 2003. Medical therapy in patients
 390 undergoing percutaneous coronary intervention: results from the ROSETTA registry. Can J
 391 Cardiol 19:1009-1015.
- 392 13. Roberts JA, Stove V, De Waele JJ, Sipinkoski B, McWhinney B, Ungerer JP, Akova M,
 393 Bassetti M, Dimopoulos G, Kaukonen KM, Koulenti D, Martin C, Montravers P, Rello J,
 394 Rhodes A, Starr T, Wallis SC, Lipman J. 2014. Variability in protein binding of teicoplanin and
 395 achievement of therapeutic drug monitoring targets in critically ill patients: Lessons from the
 396 DALI Study. Int J Antimicrob Agents 43:423-430.
- 397 14. **Sanofi.** 2016. Summary of Product Characteristics for Targocid[®].
- 398 15. Pea F, Viale P, Candoni A, Pavan F, Pagani L, Damiani D, Casini M, Furlanut M. 2004.
 399 Teicoplanin in patients with acute leukaemia and febrile neutropenia: a special population benefiting from higher dosages. Clin Pharmacokinet 43:405-415.
- 401 16. Byrne CJ, Egan S, Fennell JP, O'Byrne P, Enright H, Deasy E, Ryder SA, D'Arcy DM, McHugh
 402 J. 2015. Teicoplanin use in adult patients with haematological malignancy: Exploring
 403 relationships between dose, trough concentrations, efficacy and nephrotoxicity. Int J
 404 Antimicrob Agents 46:406-412.

- 405 17. **Brink AJ, Richards GA, Colombo G, Bortolotti F, Colombo P, Jehl F.** 2014. Multicomponent antibiotic substances produced by fermentation: implications for regulatory authorities, critically ill patients and generics. Int J Antimicrob Agents **43:**1-6.
- 408 18. **Schmidt S, Gonzalez D, Derendorf H.** 2010. Significance of protein binding in pharmacokinetics and pharmacodynamics. J Pharm Sci **99:**1107-1122.
- 410 19. Yano R, Nakamura T, Tsukamoto H, Igarashi T, Goto N, Wakiya Y, Masada M. 2007.
 411 Variability in teicoplanin protein binding and its prediction using serum albumin
 412 concentrations. Ther Drug Monit 29:399-403.
- 413 20. Mimoz O, Rolland D, Adoun M, Marchand S, Breilh D, Brumpt I, Debaene B, Couet W.
 414 2006. Steady-state trough serum and epithelial lining fluid concentrations of teicoplanin 12
 415 mg/kg per day in patients with ventilator-associated pneumonia. Intensive Care Med
 416 32:775-779.
- 417 21. Brink AJ, Richards GA, Lautenbach EE, Rapeport N, Schillack V, van Niekerk L, Lipman J,
 418 Roberts JA. 2015. Albumin concentration significantly impacts on free teicoplanin plasma
 419 concentrations in non-critically ill patients with chronic bone sepsis. Int J Antimicrob Agents
 420 45:647-651.
- 421 22. Pea F, Viale P, Furlanut M. 2005. Antimicrobial therapy in critically ill patients: a review of
 422 pathophysiological conditions responsible for altered disposition and pharmacokinetic
 423 variability. Clin Pharmacokinet 44:1009-1034.
- 424 23. Klastersky J, Paesmans M, Rubenstein EB, Boyer M, Elting L, Feld R, Gallagher J, Herrstedt
 425 J, Rapoport B, Rolston K, Talcott J. 2000. The Multinational Association for Supportive Care
 426 in Cancer risk index: A multinational scoring system for identifying low-risk febrile
 427 neutropenic cancer patients. J Clin Oncol 18:3038-3051.
- 428 24. Charlson ME, Pompei P, Ales KL, MacKenzie CR. 1987. A new method of classifying
 429 prognostic comorbidity in longitudinal studies: development and validation. J Chronic Dis
 430 40:373-383.
- 431 25. **Devine B.** 1974. Gentamicin therapy. Drug Intell Clin Pharm **8:**650-655.
- 432 26. Mosteller RD. 1987. Simplified calculation of body-surface area. N Engl J Med 317:1098.
- 433 27. **Cockcroft DW, Gault MH.** 1976. Prediction of creatinine clearance from serum creatinine. Nephron **16:**31-41.
- 435 28. **Jelliffe R.** 2002. Estimation of creatinine clearance in patients with unstable renal function, without a urine specimen. Am J Nephrol **22**:320-324.
- 437 29. Levey AS, Coresh J, Greene T, Stevens LA, Zhang YL, Hendriksen S, Kusek JW, Van Lente F.
 438 2006. Using standardized serum creatinine values in the modification of diet in renal disease study equation for estimating glomerular filtration rate. Ann Intern Med 145:247-254.
- 440 30. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, 3rd, Feldman HI, Kusek JW, Eggers
 441 P, Van Lente F, Greene T, Coresh J. 2009. A new equation to estimate glomerular filtration
 442 rate. Ann Intern Med 150:604-612.
- 443 31. Ueda T, Takesue Y, Nakajima K, Ichki K, Wada Y, Komatsu M, Tsuchida T, Takahashi Y,
 444 Ishihara M, Kimura T, Uchino M, Ikeuchi H. 2014. High-dose regimen to achieve novel target
 445 trough concentration in teicoplanin. J Infect Chemother 20:43-47.
- Niwa T, Imanishi Y, Ohmori T, Matsuura K, Murakami N, Itoh Y. 2010. Significance of individual adjustment of initial loading dosage of teicoplanin based on population pharmacokinetics. Int J Antimicrob Agents 35:507-510.

452

449 33. Yamada T, Nonaka T, Yano T, Kubota T, Egashira N, Kawashiri T, Oishi R. 2012. Simplified
 450 dosing regimens of teicoplanin for patient groups stratified by renal function and weight
 451 using Monte Carlo simulation. Int J Antimicrob Agents 40:344-348.

453	Figure Legends
454	
455	FIG 1 Teicoplanin trough total concentrations (left-hand plot) and trough unbound concentrations
456	(right-hand plot) on Days 3, 4, 7 and 10. Data are presented as median, IQR and range.
457	

458	
459	FIG 2 Relationship between trough total and trough unbound teicoplanin concentrations. The solid
460	line is a least-squares fit to the data. Pearson correlation coefficient of 0.722 (P <0.001, n =30) and
461	0.692 (P <0.001, n =29) for trough concentrations at 48 h and 72 h, respectively.
462	

FIG 3 Relationship between percentage of unbound teicoplanin and serum albumin concentration.

Trough unbound concentrations taken on Days 3, 4, 7 and 10 are included in the plot (*n*=95). The

curved line is the quadratic least-squares fit to the data. Pearson correlation coefficient of -0.599

(*P*<0.001).

FIG 4 Comparison of renal function estimation equations versus measured urinary creatinine clearance in the study population (*n*=30). The diagonal lines in each plot represent lines of x=y.

CG-TBW and CG-IBW, estimated creatinine clearance calculated by the Cockcroft-Gault equation (27) using total body weight and ideal body weight calculated by the Devine equation (25), respectively;

MDRDa, estimated glomerular filtration rate calculated by the 4-variable Modification of Diet in

Renal Disease equation (29) adjusted to the body surface area of the individual patient calculated by the Mosteller equation (26); MDRD, estimated glomerular filtration rate calculated by the 4-variable Modification of Diet in Renal Disease equation; CKD-EPI, estimated glomerular filtration rate calculated by the Chronic Kidney Disease Epidemiology Collaboration equation (30); and JEL,

estimated creatinine clearance calculated by the Jelliffe equation (28).

FIG 5 Comparison of model-predicted versus observed trough total teicoplanin concentrations in study patients using measured creatinine clearance (Measured CLcr model), and using estimated creatinine clearance, calculated by the Cockcroft-Gault equation (27) using total body weight (Estimated CLcr model), as a substitute for measured creatinine clearance. The diagonal lines in each plot represent lines of x=y.

488 Tables

489

490

TABLE 1 Demographic and clinical details of the included patients $(n=30)^a$

TABLE 1 Demographic and chinical details of the included patients (17–30)				
Male sex	14 (46.7)			
Age (years)	64 [14]			
Total body weight (kg)	69.1 ± 15.8			
Ideal body weight (kg)	56.7 ± 10.1			
Body mass index (kg/m²)	26.0 ± 5.3			
Creatinine clearance (mL/min) ^b	72 ± 41			
Serum albumin concentration (g/L) ^b	29 [4]			
Fluid input (L) ^b	2.8 ± 1.1			
Haematological malignancy diagnosis				
Acute lymphoblastic leukaemia	1 (3.3)			
Acute myeloid leukaemia	7 (23.3)			
Chronic lymphocytic leukaemia	1 (3.3)			
Hodgkin's lymphoma	1 (3.3)			
Non-Hodgkin's lymphoma	13 (43.3)			
Multiple myeloma	6 (20.0)			
Myelodysplastic syndrome	1 (3.3)			
Bone marrow transplant received	7 (23.3)			
MASCC risk-index score (23)	16 [5]			
Charlson co-morbidity index (24)	6 [3]			
Severe neutropaenia ^c	25 (83.3)			
Mean loading dose (mg/kg) ^d	9.5 ± 1.9			
Mean daily maintenance dose (mg/kg)	10.0 ± 1.8			
Duration of therapy (days)	9 ± 4			

MASCC, Multinational Association for Supportive Care in Cancer

 $^{^{}a}$ Data are presented as the mean \pm SD or the median [IQR] for continuous variables, and as the number (%) for categorical variables.

^b Value on Day 3 of teicoplanin therapy.

^c Severe neutropaenia defined as an absolute neutrophil count of <0.5 x 10⁹/L.

^d Administered for three doses at the start of teicoplanin therapy.

TABLE 2 Comparison of the performance of renal function estimation equations relative to measured creatinine clearance in the study population (n=30)

Equation	Median	IQR for	% of estimates	Root mean
	difference	differences	within 30% of	square error
	(bias) ^a	(precision) ^a	measured CL _{CR}	(accuracy)
			(accuracy)	
CG-TBW (mL/min)	3.0	29.5	63.3	29.5
CG-IBW (mL/min)	-8.5	44.3	53.3	38.9
MDRD $(mL/min/1.73 m^2)$	2.0	48.8	53.3	39.2
MDRDa (mL/min)	1.0	44.0	53.3	36.6
CKD-EPI (mL/min/1.73 m ²)	3.0	32.0	60.0	29.1
JEL $(mL/min/1.73 m^2)$	2.5	30.3	66.7	32.0

CL_{CR}, creatinine clearance; CG-TBW, Cockcroft-Gault equation (27) using total body weight; CG-IBW, Cockcroft-Gault equation using ideal body weight calculated by the Devine equation (25); MDRD, 4-variable Modification of Diet in Renal Disease equation (29); MDRDa, 4-variable Modification of Diet in Renal Disease equation adjusted to the body surface area of the individual patient calculated by the Mosteller equation (26); CKD-EPI, Chronic Kidney Disease Epidemiology Collaboration equation (30); JEL, Jelliffe equation (28). ^a Difference refers to estimated value minus measured CL_{CR}.

494

TABLE 3 Comparison of successful versus failed teicoplanin treatments for cases of coagulase-negative staphylococcal central line associated bloodstream infection $(N=7)^a$

	Success (n=4)	Failure (n=3)	<i>P</i> -value ^b
Male sex	1 (25.0)	2 (66.7)	0.486
Age (years)	57 ± 27	61 ± 12	0.796
Total body weight (kg)	54.8 ± 8.9	76.4 ± 20.5	0.230
Creatinine clearance (mL/min) ^c	50 ± 29	75 ± 13	0.195
Serum albumin concentration (g/L) ^c	30 [5]	32 [13]	0.629
Severe neutropaenia ^d	3 (75.0)	2 (66.7)	1.000
MASCC risk-index score (23)	18 [2]	16 [8]	0.057
Charlson co-morbidity index (24)	6 ± 3	6 ± 2	0.731
Mean loading dose (mg/kg)	11.2 ± 2.0	8.7 ± 0.9	0.093
Mean daily maintenance dose (mg/kg)	11.2 ± 2.0	10.9 ± 1.4	0.830
Combination therapy e	3 (75.0)	3 (100.0)	1.000
Teicoplanin MIC (mg/L) ^f	1.4 ± 1.4	1.5 ± 0.5	0.900
Trough total concentration at 48 h (mg/L)	18.6 ± 12.3	12.6 ± 7.6	0.471
Trough unbound concentration at 48 h (mg/L)	1.6 ± 1.2	1.3 ± 1.0	0.711
Trough total concentration at 72 h (mg/L)	22.8 ± 15.2	16.4 ± 5.5	0.495
Trough unbound concentration at 72 h (mg/L)	1.75 ± 1.25	1.5 ± 0.9	0.770
Trough total concentration at 48 h/MIC ^f	30.9 ± 24.4	10.3 ± 8.2	0.297
Trough unbound concentration at 48 h/MIC ^f	2.8 ± 2.2	1.1 ± 1.2	0.321
Trough total concentration at 72 h/MIC ^f	38.1 ± 29.9	12.7 ± 7.7	0.289
Trough unbound concentration at 72 h/MIC ^f	3.4 ± 2.7	1.2 ± 1.1	0.325

MASCC, Multinational Association for Supportive Care in Cancer; MIC, minimum inhibitory concentration.

^a Data are presented as the mean ± SD or the median [IQR] for continuous variables, and as the number (%) for categorical variables.

^b P-value: Fisher's exact test for categorical covariates, unpaired Student's *t*-test or Mann-Whitney *U* test for continuous covariates.

^c Values on Day 3 of teicoplanin therapy.

^d Severe neutropaenia defined as an absolute neutrophil count of <0.5 x 10⁹L⁻¹.

^e Combination therapy: other antibiotics administered concurrently with teicoplanin – piperacillin/tazobactam (4), gentamicin (3), meropenem (3), ciprofloxacin (2) and amikacin (1).

^fResult based on 3 successful treatments and 3 failures.

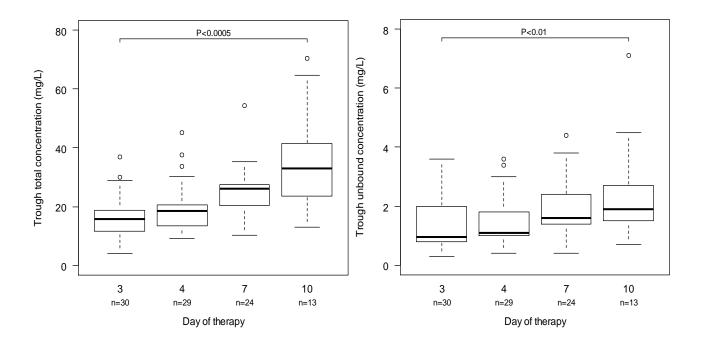


FIG 1 Teicoplanin trough total concentrations (left-hand plot) and trough unbound concentrations (right-hand plot) on Days 3, 4, 7 and 10. Data are presented as median, IQR and range.

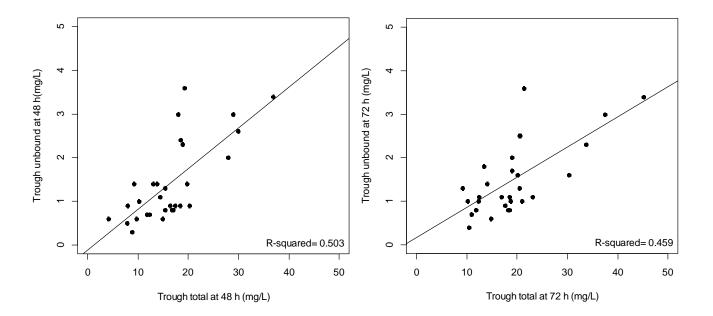


FIG 2 Relationship between trough total and trough unbound teicoplanin concentrations. The solid line is a least-squares fit to the data. Pearson correlation coefficient of 0.722 (P<0.001, n=30) and 0.692 (P<0.001, n=29) for trough concentrations at 48 h and 72 h, respectively.

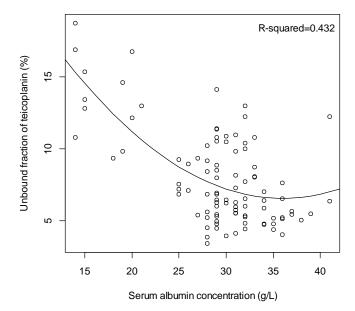


FIG 3 Relationship between percentage of unbound teicoplanin and serum albumin concentration. Trough unbound concentrations taken on Days 3, 4, 7 and 10 are included in the plot (n=95). The curved line is the quadratic least-squares fit to the data. Pearson correlation coefficient of -0.599 (P<0.001).

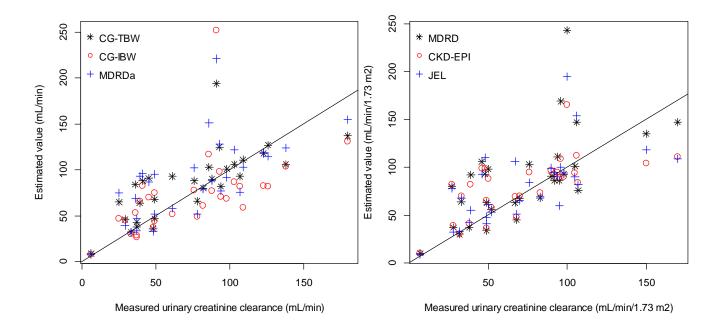


FIG 4 Comparison of renal function estimation equations versus measured urinary creatinine clearance in the study population (*n*=30). The diagonal lines in each plot represent lines of x=y. CG-TBW and CG-IBW, estimated creatinine clearance calculated by the Cockcroft-Gault equation (27) using total body weight and ideal body weight calculated by the Devine equation (25), respectively; MDRDa, estimated glomerular filtration rate calculated by the 4-variable Modification of Diet in Renal Disease equation (29) adjusted to the body surface area of the individual patient calculated by the Mosteller equation (26); MDRD, estimated glomerular filtration rate calculated by the 4-variable Modification of Diet in Renal Disease equation; CKD-EPI, estimated glomerular filtration rate calculated by the Chronic Kidney Disease Epidemiology Collaboration equation (30); and JEL, estimated creatinine clearance calculated by the Jelliffe equation (28).

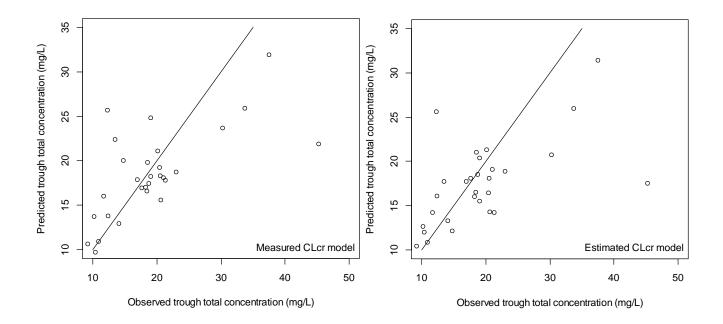


FIG 5 Comparison of model-predicted versus observed trough total teicoplanin concentrations in study patients using measured creatinine clearance (Measured CLcr model), and using estimated creatinine clearance, calculated by the Cockcroft-Gault equation (27) using total body weight (Estimated CLcr model), as a substitute for measured creatinine clearance. The diagonal lines in each plot represent lines of x=y.