

1                   **The role of macrophages in the infarcted myocardium:**  
2                                   **orchestrators of ECM remodelling**

3  
4   **Sinead A. O'Rourke<sup>1,2,3</sup>, Aisling Dunne<sup>2</sup>, Michael G. Monaghan<sup>\*1,3,4</sup>**

5   <sup>1</sup>Department of Mechanical and Manufacturing Engineering, Trinity College Dublin, Dublin,  
6   Ireland

7  
8   <sup>2</sup>School of Biochemistry & Immunology and School of Medicine, Trinity Biomedical Science  
9   Institute, Trinity College Dublin, Dublin, Ireland

10  
11   <sup>3</sup>Trinity Centre for Bioengineering, Trinity Biomedical Science Institute, Trinity College Dublin,  
12   Ireland

13  
14   <sup>4</sup>Advanced Materials for BioEngineering Research (AMBER) Centre, Trinity College Dublin  
15   and Royal College of Surgeons in Ireland, Dublin 2, Ireland.

16  
17   **\*Correspondence:** Michael Monaghan, [monaghmi@tcd.ie](mailto:monaghmi@tcd.ie)

18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32

33 **Abstract**

34 Myocardial infarction is the most common form of acute cardiac injury attributing to heart failure.  
35 While there have been significant advances in current therapies, mortality and morbidity remain  
36 high. Emphasis on inflammation and extracellular matrix remodelling as key pathological factors  
37 has brought to light new potential therapeutic targets including macrophages which are central  
38 players in the inflammatory response following myocardial infarction. Blood derived and tissue  
39 resident macrophages exhibit both a pro- and anti-inflammatory phenotype, essential for removing  
40 injured tissue and facilitating repair, respectively. Sustained activation of pro-inflammatory  
41 macrophages evokes extensive remodelling of cardiac tissue through secretion of matrix proteases  
42 and activation of myofibroblasts. As the heart continues to employ methods of remodelling and  
43 repair, a destructive cycle prevails ultimately leading to deterioration of cardiac function and heart  
44 failure. This review summarizes not only the traditionally accepted role of macrophages in the  
45 heart but also recent advances that have deepened our understanding and appreciation of this  
46 dynamic cell. We discuss the role of macrophages in normal and maladaptive matrix remodelling,  
47 as well as studies to date which have aimed to target the inflammatory response in combatting  
48 excessive matrix deposition and subsequent heart failure.

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66 **Introduction**

67 Heart failure, is a global pandemic, accounting for 31% of deaths worldwide (1). Health  
68 expenditures associated with heart failure are substantial, and expected to increase dramatically  
69 with an ageing population (2). Myocardial infarction (MI) is the most common form of acute  
70 cardiac injury attributing to heart failure, and while there have been significant advances in  
71 therapies, mortality and morbidity remain high. Our understanding of MI has evolved in recent  
72 years with inflammation driven by macrophages now recognised as playing a key pathological role  
73 in the progression of tissue remodelling and fibrosis which, in turn, limits cardiac function. A  
74 greater appreciation of the role of the inflammatory response and the interaction between  
75 macrophages and the extracellular matrix (ECM) is required in order to provide greater insight  
76 into tissue remodelling and disease progression within the myocardium, as well as revealing  
77 therapeutic targets for the treatment of heart failure. In this review we will discuss the importance  
78 and role of macrophages in the healthy and infarcted myocardium, and how these innate immune  
79 cells contribute towards ECM remodelling and fibrosis.

80

81 **Multicellularity of the heart**

82 The myocardium is a multicellular complex tissue comprised of a range of distinct cell-types.  
83 Cardiomyocytes (CMs) constitute approximately one third of resident myocardial cells by number  
84 (3), with the remaining two thirds referred to as non-excitabile cells (non-CMs) such as fibroblasts,  
85 smooth muscle cells, endothelial cells, autonomic motor neurons, and immune cells such as mast  
86 cells and macrophages (4). While CMs possess inherent conduction capabilities which mediate the  
87 characteristic contractile forces of the heart, non-CMs are responsible for matrix deposition,  
88 vascularization and autonomic regulation (5). CMs and non-CMs communicate via biochemical  
89 signalling through cytokine and growth factor secretion (5, 6). Such signals arise, for example,  
90 during development and regulation and include the release of vascular endothelial growth factor  
91 (VEGF) which activates endothelial cells to initiate angiogenesis (7), or in response to trauma or  
92 injury, where signalling is mediated by pro-inflammatory cytokines such tumour necrosis factor  
93 alpha, (TNF $\alpha$ ) (8, 9). Numerous networks also exist between non-CMs such as fibroblasts and  
94 macrophages, working in tangent to maintain the structural integrity of the heart.

95 Fibroblasts are traditionally defined as cells of mesenchymal origin, arising from bone marrow  
96 derived cells known as fibrocytes (10, 11). Cardiac fibroblasts produce the necessary components  
97 for the construction of the ECM in order to maintain the integrity of the myocardium (10). As a  
98 result, fibroblasts have been highlighted as key mediators of both normal cardiac function and the  
99 remodelling response to injury (6, 10, 12). In addition to producing components of the ECM,  
100 fibroblasts are also observed to secrete regulatory proteins and matrix metalloproteinases as well  
101 as their corresponding inhibitors (tissue inhibitors of metalloproteinases), thus maintaining a well-  
102 controlled balance for ECM homeostasis (13).

103

104

105

106 **Macrophages – key drivers of the innate immune response**

107 Macrophages (and their precursors, monocytes) are key mediators of the innate immune response  
108 involved in the recognition, phagocytosis and elimination of pathogens. They exist as both  
109 circulating and tissue resident cells within the body and have the ability to change their function  
110 and phenotype based on environmental cues(14). While they exist as a heterogenous population  
111 they can be broadly classified as M1 or M2 macrophages (15). M1 macrophages are traditionally  
112 associated with a pro-inflammatory response, and are referred to as classically activated  
113 macrophages, induced by IFN $\gamma$ , LPS, and TNF $\alpha$ . When stimulated, M1 macrophages secrete high  
114 levels of pro-inflammatory cytokines IL-12, IL-23, IL-1 and IL-6 (16). M2 macrophages, or  
115 “alternatively activated” macrophages exhibit an anti-inflammatory, pro-regenerative phenotype  
116 largely due to their ability to secrete high levels of anti-inflammatory cytokines including IL-10  
117 and growth factors such as VEGF as well as matrix metalloproteinases (MMPs) (16). In murine  
118 models, M1 and M2 macrophages are distinguished from another through the expression of the  
119 inflammatory monocyte marker Ly6C. Ly6C<sup>high</sup> monocytes are preferentially recruited to sites of  
120 inflammation and exhibit an M1 pro-inflammatory phenotype while Ly6C<sup>low</sup> monocytes represent  
121 the non-classical population and differentiate into M2 macrophages to promote tissue healing and  
122 angiogenesis(17).

123 While the M1/M2 paradigm proves useful as a preliminary introduction to these innate immune  
124 cells, the full story is not as black and white. The macrophage phenotype exhibits more plasticity  
125 than historically assumed, and M1/M2 classification merely represents two extremes of a  
126 continuum of activated states. For example, macrophages treated with the pathogen associated  
127 molecule, lipopolysaccharide (LPS), exhibit a reduced phagocytic capacity compared to  
128 macrophages treated with the endogenous cytokine; IFN $\gamma$ . While both produce pro-inflammatory  
129 mediators, LPS polarized macrophages are now referred to as M1b macrophages, while IFN $\gamma$   
130 polarized macrophages are referred to as M1a macrophages. (16, 18).

131 Distinct M2 macrophage subsets also exist. For example, M2a macrophages are induced by IL-4  
132 and IL-13 and have a pre-dominantly anti-inflammatory phenotype, secreting high levels of IL-10  
133 and IL-1 receptor antagonist as a means of dampening the inflammatory response. M2b  
134 macrophages, on the other hand, exhibit both pro and anti-inflammatory responses, producing IL-  
135 1 $\beta$ , TNF $\alpha$  and IL-6 as well as IL-10 in response to LPS stimulation. M2c macrophages are induced  
136 by IL-10 and secrete high levels of TGF $\beta$ 1 (transforming growth factor beta 1) and glucocorticoids.  
137 They assume a regenerative, pro-healing phenotype and play a major role in promoting tissue  
138 repair and silencing the inflammatory response. These cells also play a significant role in matrix  
139 deposition (15). More recently, M2d and M2f phenotypes have been characterised (19, 20). M2d  
140 macrophages are activated by Toll-like receptor agonists and adenosine A2a receptor agonists. In  
141 response, these cells secrete high levels of VEGF and IL-10, and in turn downregulate TNF $\alpha$  and  
142 IL-12 production (19). M2f cells are induced by efferocytosis which involves the removal of  
143 apoptotic cells by macrophages. This process is similar to phagocytosis, however, it involves  
144 distinct receptors and signalling pathways and results in the secretion of high levels of TGF $\beta$ 1,  
145 prostaglandin E2 and platelet activating factor, all of which are known to inhibit LPS-induced pro-  
146 inflammatory cytokine production(21)

147 **Tissue Resident Macrophages**

148 Tissue resident macrophages exist at various sites throughout the body and can include microglia  
149 in the brain and Kupffer cells in the liver (22). The heart, being no exception; contains its own  
150 resident macrophages which possess a specific role in the regulation of cardiac function (23). The  
151 distinction between tissue residing cardiac macrophages and circulating monocyte-derived  
152 macrophages has become a considerable area of focus in recent years (24). While long established  
153 tissue resident cells appear to facilitate coronary development and tissue homeostasis, it appears  
154 that monocyte-derived infiltrating cells have a predominant role in tissue injury and destruction.  
155 This highlights that macrophages, whether circulating or permanently residing, originate from  
156 diverse lineages, and as a result have different functions.

157

158 **CCR2<sup>+</sup> and CCR2<sup>-</sup> tissue resident macrophages**

159 Gene mapping of cardiac resident macrophages reveals two distinct lineages arising at the  
160 embryonic stage and postnatal stage (25). Developmental studies of early cell migration in murine  
161 models affirms this, with the earliest cardiac resident macrophages derived from an erythromyeloid  
162 progenitor in the yolk sac (26). These progenitor macrophages migrate out of the yolk-sac either  
163 directly to the developing myocardium or else to the foetal liver, where they progress to  
164 haemopoietic stem cells and eventually cardiac tissue-resident macrophages (26). Postnatally,  
165 monocyte-derived macrophages can also migrate to the myocardium to become tissue resident  
166 macrophages.(27). These embryonic and postnatal resident cells can be distinguished from one  
167 another based on expression of the chemokine receptor, Chemokine Receptor Type 2 (CCR2)(25).  
168 This receptor and its corresponding ligand, chemokine ligand 2 (CCL2), also known as monocyte  
169 chemoattractant protein 1 (MCP-1), play an important role in monocyte/macrophage migration.  
170 Studies have demonstrated that CCR2<sup>+</sup> cardiac resident macrophages are derived from monocytes  
171 while CCR2<sup>-</sup> macrophages originate from the embryonic developmental stage (25, 28).  
172 Furthermore, CCR2<sup>-</sup> macrophages undergo local proliferation in order to replenish their population  
173 whereas CCR2<sup>+</sup> macrophages are repopulated by monocyte-derived macrophages extravasating  
174 into the myocardium (25). Both CCR2<sup>+</sup> and CCR2<sup>-</sup> cell populations orchestrate diverse responses  
175 following traumatic events such as MI. CCR2<sup>+</sup> cells facilitate monocyte recruitment into the heart  
176 following MI via CCR2-MCP1 mediated trafficking and secrete high levels of pro-inflammatory  
177 mediators including IL-1 $\beta$ , TNF and IL-6 (28). Not surprisingly, depletion of this cell population  
178 has resulted in reduced infarct size in a murine model of MI (28).

179 Conversely, CCR2<sup>-</sup> macrophages appear to play an immune-modulatory, pro-regenerative role,  
180 expressing high levels of growth factors including Insulin-like growth factor 1 (IGF1) and Platelet  
181 derived growth factor C (PDGF-C) (28). Depletion of this CCR2<sup>-</sup> population has enhanced  
182 monocyte/macrophage infiltration to the heart and further implicates these cells as potential  
183 immune-modulators during MI (28, 29). In addition to immune modulatory functions, recent  
184 studies have also demonstrated that CCR2<sup>-</sup> macrophages express high levels of the sodium  
185 channel, SCN4, and sodium channel modulator, FGF13, suggesting that macrophages can  
186 modulate the electrical activity of cardiomyocytes (25, 30). Fracktalkine receptor (CX3CR1<sup>+</sup>)  
187 expressing resident macrophages have also been reported to facilitate conductivity, further

188 implicating their role in regular functioning of the heart and broadening the role of macrophages  
189 beyond local inflammation and immune-modulation (30). It is not yet clear if both CCR2<sup>+</sup> and  
190 CCR2<sup>-</sup> macrophages contribute to the electrical homeostasis of the heart and, given that both  
191 subsets express the CX3CR1 (31), delineation of the direct impact of these individual cell-types  
192 on cellular conductivity is a promising avenue of exploration.

193

### 194 **The Extracellular Matrix**

195 The ECM is a complex and dynamic structure of hundreds of numerous proteins which provide a  
196 support system for cells. Within the myocardium, it acts as a mechanical scaffold to create cellular  
197 and acellular networks. Conceptually, the cardiac ECM can be divided into two segments, the  
198 interstitial matrix, comprised of primarily type I and type III collagen, and the basement  
199 membrane, comprised of collagen IV, V, VII, X as well as laminins (32). Proteins residing in the  
200 interstitial matrix and basement membrane of the ECM serve a specific function, either facilitating  
201 structural support of the matrix itself, or regulating local cell behaviour and function (33). Type I  
202 and Type III collagens for example allow the myocardium to maintain structural integrity through  
203 tensile support. Cardiac tissue undergoes mechanical stress via shear and pressure forces of muscle  
204 contraction and the organization and continuity of the collagen networks within the ECM allows  
205 for appropriate distribution of this physical stress. Elastin in the interstitial matrix provides  
206 elasticity to the cardiac tissue, with reduced expression post MI contributing to stiffer scar tissue  
207 (34). Proteoglycans along with glycoproteins play a key role in signalling and turnover of the ECM  
208 (35), highlighting the alternative function of ECM as a transducer of signals within the cardiac  
209 environment.

210 The ECM is not an inert structure, with matrix continuously responding to signals from the  
211 surrounding environment and exerting its own signalling through mechanical and chemical cues.  
212 In the context of MI and chronic inflammation, disruption to the ECM via adverse remodelling  
213 leads to disarray of physical stress, applying strain on the myocardium and leading to dysfunction.  
214 Key initiators of remodelling can include ischaemia, pressure overload, and ageing of the heart,  
215 all of which have significant association with systemic inflammation (36-38). Thus, it is  
216 established that remodelling of the ECM as a consequence of sustained inflammation, is a critical  
217 etiological factor of heart failure, making the study of ECM and the innate immune activity in the  
218 failing myocardium one of great importance (39, 40).

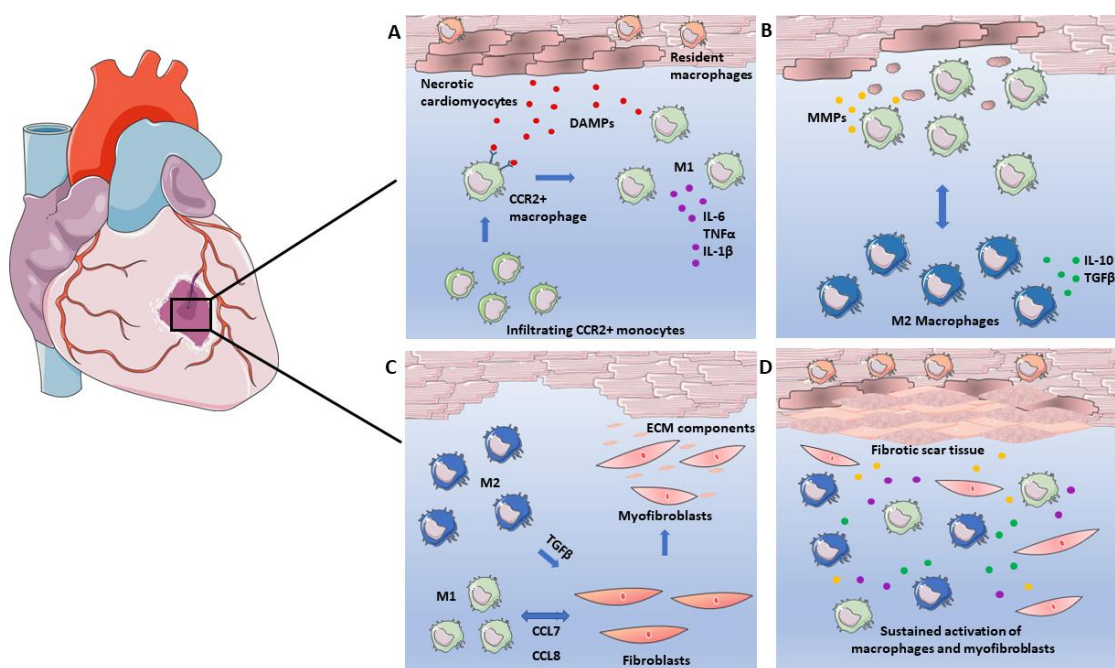
219

### 220 **Role of inflammation in heart failure -MI and ischaemia**

221 MI refers to mass cardiomyocyte death as a result of ischemia, which is often worsened by a  
222 subsequent reperfusion of oxygen supply, and the ensuing inflammatory response (41). This  
223 association between inflammation and adverse cardiac events is well acknowledged. Multiple  
224 studies have demonstrated that elevated pro-inflammatory cytokine production in the heart  
225 correlates with worsening outcome. Furthermore, inhibition of pro-inflammatory cytokines such  
226 as TNF $\alpha$ , which is heavily implicated in cardiac disease, results in improved cardiac function in  
227 rat models of heart failure (8, 9). Two classifications of infarction are often presented, both which

228 occur as a result of ischemia. Acute MI is caused by an atherosclerotic plaque rupture causing  
 229 coronary artery occlusion and cardiac tissue damage due to ischemia. Chronic MI refers to  
 230 continued loss of cardiomyocytes from gradual and prolonged ischemia, often greater than 8  
 231 weeks. The vast amount of cell death following MI poses a detriment, as the heart itself possesses  
 232 a limited regenerative capacity (42). Left untreated, cardiac tissue undergoes extensive  
 233 remodelling to compensate for cell loss and to maintain structural integrity. The inflammatory  
 234 response facilitates the removal of necrotic cells in addition to tissue remodelling (43). However,  
 235 extensive remodelling imposes stress on the heart, instigating maladaptive mechanisms such as  
 236 chronic inflammation and cellular apoptosis. As the heart continues to employ methods of  
 237 remodelling and repair to resolve this, a destructive cycle prevails, ultimately leading to  
 238 deterioration of cardiac function and heart failure (44). These events are summarised in Figure 1.

239



240

241 **Figure 1: Macrophages in the response to infarction.** A) Cardiomyocytes undergo necrosis,  
 242 releasing DAMPs and attracting CCR2<sup>+</sup> circulating monocytes. CCR2<sup>+</sup> monocytes differentiate  
 243 into pro-inflammatory M1 macrophages replacing resident macrophages and secreting high levels  
 244 of pro-inflammatory cytokines IL-6, TNF $\alpha$ , and IL-1 $\beta$ . B) M1 macrophages clear necrotic cell  
 245 debris through phagocytosis and induce breakdown of the ECM through secretion of MMPs.  
 246 Phagocytosis of the necrotic debris causes macrophage polarisation to the M2 phenotype. M2  
 247 macrophages secrete high levels of anti-inflammatory cytokine IL-10 and growth factor TGF $\beta$ . C)  
 248 Both M1 and M2 macrophages facilitate the fibrotic response. M1 macrophages recruit fibroblasts  
 249 via CCL7 and CCL8 mediated signalling. M2 macrophages induce fibroblast differentiation into  
 250 myofibroblasts, which in turn secrete ECM components to facilitate tissue repair. D) Sustained  
 251 activation of macrophages leads to continuous secretion of growth factors, pro-inflammatory  
 252 cytokines, and MMPs. Continued breakdown of ECM as well as overproduction of ECM  
 253 components by myofibroblasts leads to adverse remodelling of ECM and results in fibrotic scar  
 254 tissue.

255 Infiltration of monocyte-derived macrophages to the infarct is a key feature of MI, and can be  
256 characterised by stages of macrophage infiltration, and their subsequent actions within the  
257 myocardium. Immediately following infarction, resident cardiac macrophages begin to die in  
258 response to ischemia, with a complete loss of resident macrophages within the infarct observed in  
259 murine models 24 hours post infarction (45). The resident macrophage population lost in the  
260 ischemic region is rapidly replaced by infiltrating monocyte-derived macrophages within 24 hours  
261 (45). Day 1-3 post MI, infiltrating macrophages exhibit a pro-inflammatory M1-like phenotype,  
262 driving acute inflammation and facilitating clearance of dead cells. At approximately day 5-7 post  
263 MI, these macrophages begin to adopt a reparative M2-like phenotype, working to resolve  
264 inflammation and rebuild cardiac tissue (46). Mouse models of MI have revealed distinct subsets  
265 of infiltrating monocyte-derived macrophages, with earlier recruitment of pro-inflammatory  
266 Ly6C<sup>high</sup> macrophages dependent on CCR2/CCL2 signalling, and the later pro-regenerative  
267 Ly6C<sup>low</sup> macrophages recruited via CX3CR1 signalling (47). One can, therefore, hypothesise that  
268 inflammation and resolution is achieved in the myocardium through differential recruitment of  
269 macrophages.

270 It has also recently been demonstrated that infiltrating macrophages in murine mouse models of  
271 MI undergo metabolic reprogramming to increase oxidative phosphorylation at approximately day  
272 5 post MI (48). Increased oxidative phosphorylation in addition to fatty acid synthesis and  
273 oxidation, is a signature of the M2 phenotype (49) and implies that there is a phenotypic switch  
274 from the early pro-inflammatory state to a more pro-regenerative one. Thus, not only is there the  
275 possibility of recruitment of separate subsets of macrophages via CCR2 and CX3CR1 dependent  
276 signalling, but in addition, there is a switch from the M1 to the M2f phenotype subset based on  
277 fatty acid synthesis and oxidation. This switch is key in the appropriate resolution of inflammation  
278 and progression to a pro-healing state and is promoted through efferocytosis of cell debris at the  
279 site of injury (50). Engulfment of dead cells by macrophages has been observed to elevate fatty  
280 acid synthesis and triggers production of the anti-inflammatory, pro-reparative cytokine  
281 TGFβ1(51, 52). However, in circumstances of chronic ischaemia or severe infarction, continuous  
282 cardiomyocyte death leads to sustained activation of M1 macrophages, which have a diminished  
283 efferocytotic ability (53). This is heightened in patients suffering from diabetes and obesity  
284 whereby underlying chronic inflammation exacerbates the pro-inflammatory response to  
285 infarction. In this instance, elevated levels of the pro-inflammatory cytokines, TNFα and IL-6, as  
286 well as CRP are associated with worse patient outcome (54, 55).

287 Failure to clear dying cells also leads to the release of damage-associated molecular patterns  
288 (DAMPs) which contribute to a robust secretion of pro-inflammatory cytokines, prolonging  
289 activation of the inflammatory response and escalating damage in the myocardium. A timely  
290 switch from low efferocytotic M1 macrophages to highly efferocytotic anti-inflammatory M2  
291 macrophages is therefore necessary to clear dead cells and promote tissue repair.

292

### 293 **Alterations to the Extracellular Matrix**

294 In any circumstances of injury within the body, ECM degradation is necessary to allow for repair  
295 and/or replacement of the damaged tissue. In the instance of MI, ECM degradation is triggered as



296 early as ten minutes following an ischemic event (56). Matrix metalloproteinases (MMPs),  
297 predominantly produced by macrophages and fibroblasts, are secreted as part of a programmed  
298 inflammatory response in order to deconstruct matrix architecture. As MI progresses, subsequent  
299 necrosis of cardiomyocytes accentuates matrix degradation. MMPs target various components of  
300 the ECM, for example, the collagenases MMP1, MMP8 and MMP13 cleave the  $\alpha$ -chains of type  
301 I and type II collagens while MMP3 and MMP10 target proteoglycans, fibronectin and laminins  
302 (33). Gelatinases such as MMP2 along with MMP9 digest gelatin in addition to degrading type IV  
303 collagen, the most abundant component of the basement membrane (57).

304 As ECM breakdown persists, a provisional matrix enriched in fibrins forms in its place (58). This  
305 plasma-derived matrix does not serve a particularly structural role in the same manner as native  
306 ECM, but rather modulates cell phenotype and behaviour, setting the stage for tissue repair.  
307 Components of this provisional plasma-derived matrix interact with migrating cells such as  
308 macrophages via cell surface integrins. It has been hypothesised that the provisional matrix is  
309 capable of modulating gene expression and immune cell phenotype through these integrin-  
310 mediated interactions and, thus, progress the repair of cardiac tissue (59). Furthermore, the  
311 provisional matrix acts as a reservoir for numerous growth factors including PDGF, VEGF and  
312 members of the TGF family (60). These growth factors are secreted by pro-regenerative cells such  
313 as M2 macrophages and subsequently deposited within the provisional matrix, bound via the  
314 heparin binding domain (60). Sequestering of growth factors in this fashion regulates their function  
315 and may influence the activation of fibroblasts and vascular cells. However, the full extent of these  
316 processes remains to be understood.

317 Healing of the infarct area greatly relies on clearance of this provisional matrix which has been  
318 clearly demonstrated in mouse models. In mice lacking the plasminogen/plasmin system  
319 responsible for this clearance, a lack of leukocyte infiltration into the infarct region is observed,  
320 and thus repair of the myocardium impeded (61). Fragments of the provisional matrix are  
321 endocytosed by CCR2+ macrophages and postulated to induce a switch to an anti-inflammatory,  
322 reparative state within the infarct area (62). *In vitro*, interactions with a fibrin matrix lead bone  
323 marrow derived macrophages to adopt an M2 like anti-inflammatory state (63), therefore it has  
324 been suggested that the fibrin enriched matrix and its removal promote an anti-inflammatory  
325 phenotype in local macrophages. Removal of the provisional matrix is followed by secretion of  
326 cellular fibronectin to form a cell-derived matrix(33). This matrix is enriched with  
327 macromolecules, such as thrombospondins, which facilitate the recruitment and activation of  
328 fibroblasts and macrophages to a pro-regenerative phenotype in order to promote healing.

329 The dynamics of the ECM, from native, to plasma derived, and then a cell-derived remodelled  
330 matrix, is a highly ordered process to allow for efficient transition from the inflammatory response  
331 to wound healing. Any anomalies in this process can lead to sustained inflammation and fibrosis  
332 i.e. the accumulation of interstitial matrix components, predominantly collagen type I. Newly  
333 synthesised ECM differs from that of the original native ECM, with turnover of cross-linked  
334 collagen being significantly faster than that of normal collagen (64). This leads much stiffer  
335 collagen fibres, and ultimately, a stiff scar tissue post MI. High expression of lysyl oxidase  
336 (indicative of cross-linking) has been observed in murine models of infarction, and correlates with

337 a stiffer myocardium (65). In rat models of infarction, a fivefold increase of lysyl oxidase is  
338 observed at day 3, and remains elevated at day 7 post infarction (66). While the formation of scar  
339 tissue post MI is important for maintaining structural integrity while the myocardium is under  
340 reconstruction, extensive scarring or remodelling limits the functional capacity of the heart by  
341 impeding ventricular contraction and relaxation(67). Furthermore, the detrimental effects of ECM  
342 remodelling extend beyond the infarct site as formation of scar peripheral to the site of infarction  
343 is also observed. This further limits myocardial function and heightens the progression of heart  
344 failure. To greater understand how these adverse effects arise, we look to the key mediators of  
345 ECM turnover.

346

### 347 **Macrophages as mediators of ECM remodelling**

348 The cascade of events which lead to tissue remodelling post infarction may be attributed to chronic  
349 inflammation and sustained activity of pro-inflammatory macrophages within the infarcted  
350 myocardium. In the early stages of infarction (Day 0-3), the mass influx of pro-inflammatory  
351 macrophages promotes clearance of matrix and debris through phagocytosis of dying cells and  
352 secretion of matrix proteases. The secretion of pro-inflammatory cytokines including TNF $\alpha$  and  
353 IL-6 by macrophages activates resident fibroblasts, which further increases the production of  
354 MMPS such as MMP2 and MMP9(68). These “immune-activated” fibroblasts also secrete pro-  
355 inflammatory cytokines including IL-1 $\beta$  and IL-6 in response to the macrophage secretome (69),  
356 which serves as a positive feedback and augments the pro-inflammatory response. While MMP  
357 production is required for natural matrix turnover, sustained activation of pro-inflammatory  
358 macrophages, and therefore continuous production of MMPs, results in extensive matrix  
359 remodelling and impaired wound healing. High levels of MMP9 have been reported in patients,  
360 serving as a biomarker for adverse left ventricle remodelling and heart failure(70). Mice  
361 overexpressing MMP14 also show lower survival and ejection fraction following MI(71). TIMPs  
362 can also contribute to adverse remodelling if produced in abundance as this can lead to unrestricted  
363 matrix deposition, thus highlighting the need for a controlled balance between MMPs and their  
364 inhibitors(72).

365 While the influx of pro-inflammatory macrophages enhances matrix breakdown post MI, it is the  
366 transition from acute inflammation to fibrosis, facilitated by the switch from M1 to M2 dominant  
367 macrophage subsets that further exacerbates ECM remodelling. M2 macrophages secrete high  
368 levels of TGF $\beta$ 1, which drives transcription of alpha smooth muscle actin ( $\alpha$ -SMA) in the resident  
369 fibroblasts (73). As a result, these fibroblasts undergo a dramatic phenotypic transformation to  
370 become myofibroblasts(74, 75). Myofibroblasts have superior mobility compared to the  
371 homeostatic fibroblast and possess a higher capacity to produce matrix components(76).  
372 Macrophages not only amplify activation of these cells, but also facilitate their recruitment to the  
373 site of injury via signalling mediated by chemokines such as CCL7 and CCL8(77). As a result,  
374 overproduction of ECM components is observed, with an increased deposition of collagen, which  
375 stabilizes and crosslinks to form scar tissue. M2 macrophages can further promote fibrogenesis  
376 through the production of arginase, which activates glutamate and proline, both of which are  
377 necessary for collagen synthesis (78).

378 Clearly, both pro- and anti-inflammatory macrophages play a distinct pathological role in ECM  
379 remodelling, yet both subsets also have necessary roles in natural healing and repair. Therefore, it  
380 is difficult to pinpoint precisely which subset is a therapeutic target without further delineation of  
381 their functions in the infarcted myocardium. Certain pro-inflammatory cytokines, such as IL-6,  
382 elicit cardio-protective effects in the short term, and only pose a danger when their presence is  
383 sustained long-term(79-81). Furthermore, without CCR2<sup>+</sup> pro-inflammatory macrophages, the  
384 clearance of fibrin-derived provisional matrix is impaired, thus limiting progression to the more  
385 permanent cell derived matrix (62). Moreover, a prolonged presence of fibrin fragments can  
386 prompt a pro-inflammatory response (59) and contribute to a state of chronic inflammation.  
387 Conversely, eliminating M2 macrophages can lead to a worsened outcome, as they are a potent  
388 source of IL-10. This anti-inflammatory cytokine exerts protection against cardiac fibrosis, with  
389 knockout murine models demonstrating that a lack of IL-10 leads to adverse tissue remodelling  
390 and more severe cardiac fibrosis when compared to wildtype counterparts (82). A more pragmatic  
391 approach, therefore, may be to harness the effects of the macrophages through immunomodulation  
392 rather than selective elimination.

393

#### 394 **Immunomodulation: targeted therapy of heart failure**

395 While clinical studies in MI patients are limited, strategies aimed at targeting dysregulated immune  
396 responses have been explored as treatment options for heart failure post MI. Early trials involved  
397 the use of general immunosuppressants based on the hypothesis that nonspecific inflammation  
398 following MI is unfavourable (83). Such trials included a broad range of candidates that are  
399 considered the gold standard of immunosuppression such as corticosteroids, methotrexate and  
400 cyclosporin A to name but a few. However, their use in the context of cardiac treatment has yielded  
401 conflicting results. A review of clinical trials dating 1964 to 1989 reported that while  
402 corticosteroids appear to reduce mortality rates compared to placebo treatments, overall they do  
403 not provide a significant cardio-protective effect (83). Methotrexate, a well-established  
404 immunosuppressive routinely used for the treatment of rheumatoid arthritis, has also produced  
405 conflicting results with one trial reporting a reduction in TNF $\alpha$  and IL-6 together with an increase  
406 in IL-10 (84), while others reported no beneficial effects, instead worsening of left ventricle  
407 ejection fraction following treatment (85, 86). Cyclosporin A has also been considered for post-  
408 MI treatment given its ability to inhibit the mitochondrial permeability transition pore and  
409 therefore prevent necrotic cell death (87). However, no improvement of infarct size or mortality  
410 was observed in patients in a 3 day follow up (88).

411 While the lack of success for these drug trials may be attributed to short follow up periods and  
412 small sample numbers, it may also be that non-specific suppression of inflammation is insufficient  
413 to alleviate the adverse effects associated with infarction and more targeted approaches are  
414 required. IL-1 $\beta$ , for example, has proved to be a promising candidate to target due to elevated  
415 levels of the cytokine associated with poor prognosis in MI patients. Numerous pre-clinical studies  
416 have reported that inhibition of this cytokine results in reduced inflammation and remodelling in  
417 mice post infarction (89, 90). Anakinra, an established antagonist for the IL-1 $\beta$  receptor; has been  
418 assessed in multiple pilot studies for efficacy in reducing left ventricular remodelling in patients

419 (91-93). Antagonising IL-1 $\beta$  in the above studies appears to blunt the acute inflammatory response  
420 exhibited post-infarction with an increase in pro-inflammatory marker C Reactive Protein (CRP),  
421 observed following discontinuation of treatment. While the results of the 2010 study demonstrated  
422 an overall improvement in left ventricular remodelling, the small sample size proved limiting in  
423 their results. A larger study conducted in 2015 failed to show any improvement in remodelling  
424 compared to placebo treatment (92). Targeting IL-1 $\beta$  also reduces the risk of new MI events in  
425 patients with previous history of infarction. The CANTOS trial involving 10,061 patients, reported  
426 a 15% reduction in major adverse cardiovascular events upon treatment with the IL-1 $\beta$  targeting  
427 drug, canakinumab. However, no significant reduction in risk of cardiovascular death or overall  
428 mortality was observed (94).

429 Clinical trials targeting TNF $\alpha$  have also been conducted. In patients with acute MI, treatment with  
430 etanercept, a high affinity TNF receptor antagonist, resulted in reduced levels of IL-6 and lower  
431 neutrophil counts, however, no improvements in ventricular dilation or cardiac function were  
432 observed (95). Furthermore, in patients with chronic heart failure, trials involving anti-TNF $\alpha$   
433 treatment were terminated prematurely due to lack of benefit (96). Despite encouraging preclinical  
434 results from *in vivo* models, targeting single cytokines alone may not be enough to counteract the  
435 complex pathophysiology associated with heart failure, and instead, targeting the source of  
436 inflammation may represent a more viable approach.

437

### 438 ***Targeting Macrophages***

439 Targeting macrophage infiltration to combat inflammation is not an entirely new concept, however  
440 many studies have failed to show a clear efficacy *in vivo*. Previous work using small molecule  
441 inhibitors to target the migration of CCR2<sup>+</sup> macrophages, while showing a promising *in vitro*  
442 result, have failed to overcome challenges *in vivo* due to lack of tissue selectivity for the CCR2  
443 receptor as well as poor potency in administered treatments (97). Advances in short interfering  
444 RNA (siRNA) technology, including improved specificity of targeting sequences, as well as new  
445 methods of delivery, have opened the door to novel therapies to treat inflammation and heart  
446 failure. For example, siRNA targeting of the cell adhesion molecules ICAM 1/2, VCAM and E  
447 and P selectins have been shown to reduce inflammation and infarct size in a murine model of MI;  
448 ultimately preserving left ventricle ejection fraction and improving recovery after infarction (98).  
449 While this study emphasizes that a multi-targeted strategy may be necessary, targeting the CCR2  
450 receptor alone has also yielded promising results with two separate studies demonstrating that  
451 siRNA-mediated targeting of the CCR2 receptor significantly reduces infarct size in mouse models  
452 (99, 100). Specifically, siRNA targeting the CCR2 receptor 1-hour post-infarction (induced by  
453 coronary ligation) resulted in a 34% reduction in infarct size/area-at-risk at 24 hours post siRNA  
454 delivery(99). A similar study resulted in a reduction in expression of pro-inflammatory cytokines,  
455 IL-6, IL-1 $\beta$ , and TNF $\alpha$  at day 4 post infarction, while levels of IL-10 appeared to increase (100).  
456 At three weeks post infarction; a significant reduction of ventricular remodelling was observed  
457 compared to untreated mice (100). These results not only strongly implicate macrophages in the  
458 aetiology of heart failure, but also demonstrate the ability to diminish their effects through single  
459 molecule targeting, which if tissue specific, may represent a viable option for future therapy.

460 Targeting the CCR2 receptor proves particularly advantageous compared to pre-existing  
461 immunosuppressive treatments as the strategy does not affect the resident homeostatic  
462 macrophages present within the myocardium, nor does it hinder clearance of necrotic matter in the  
463 infarct. To improve siRNA delivery, nanoparticles and scaffolds are being extensively explored.  
464 Scaffolds can also be placed directly at the intended location of therapy. In particular scaffolds can  
465 also enable a controlled rate of delivery through interactions with the siRNA and designated target,  
466 as well as timed degradation of the scaffold itself (101, 102). This proves optimal for MI treatment  
467 whereby timing of inflammatory resolution is critical. Premature intervention of the inflammatory  
468 response may hinder wound repair, whereas a delayed response could fail to prevent adverse  
469 cardiac remodelling and heart failure(103).

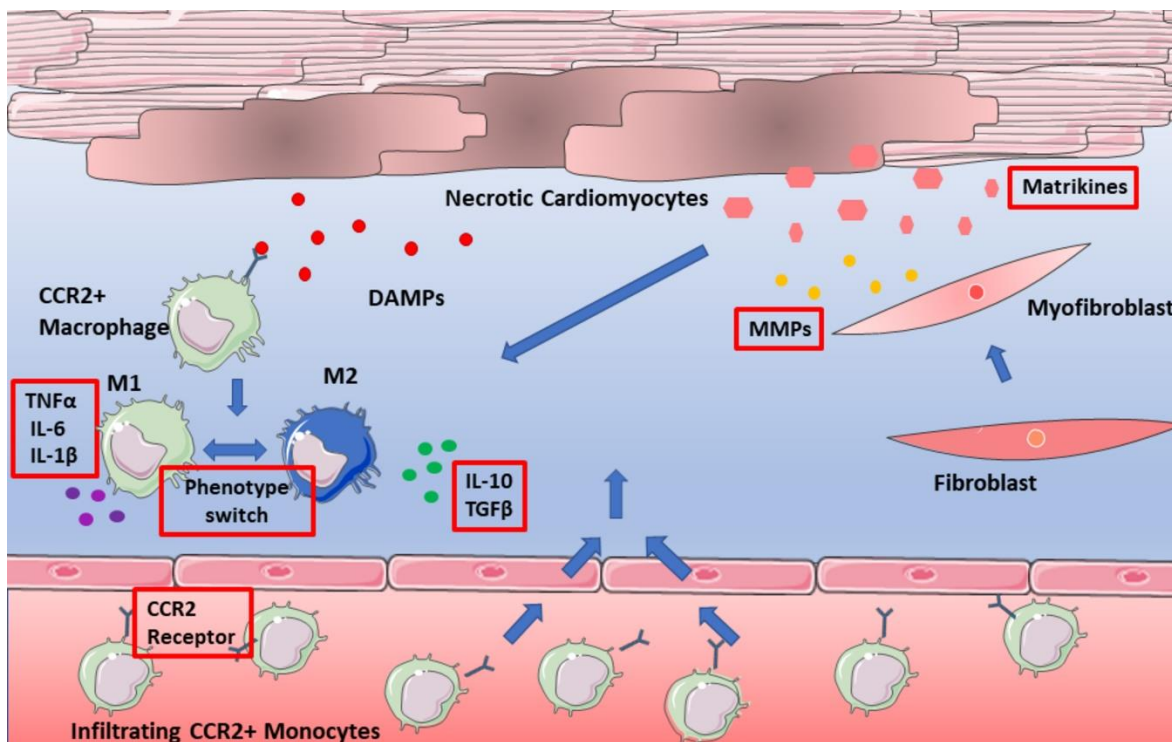
470

### 471 ***Targeting the ECM***

472 Given that the ECM plays a pivotal role in driving macrophages activation, consideration for novel  
473 therapies should also be given to the interactions between cellular matrix and macrophage. For  
474 example, the role of the metalloproteinases is not limited to breakdown of ECM components; such  
475 enzymes also have a role in regulation of the inflammatory response through proteolytic cleavage  
476 of cytokines, chemokines, and growth factors (104). Protease mediated fragmentation of matrix  
477 proteins results in the generation of ECM-derived macromolecules known as matrikines which  
478 possess a distinct role in regulation of cell activity (105). Elastin fragments and collagen peptides  
479 are the most well studied in this context and both have been implicated in numerous reports as  
480 activators of immune cells and fibroblasts during pro-inflammatory responses (106). Proline-  
481 glycine-proline, a tripeptide derived from collagen has been observed to act as a chemoattractant  
482 for neutrophil infiltration in models of pulmonary inflammation, in addition to promoting overall  
483 wound healing in mouse models. This peptide signals via the CCR2 chemokine receptor, which as  
484 mentioned previously, plays a prominent role in macrophage infiltration (107, 108). The  
485 dysregulated accumulation of these matrikines via continuous breakdown and remodelling of the  
486 ECM therefore may prove a detriment to the myocardium. Taking this into consideration, it is  
487 possible that the release of ECM fragments as well as their producers, the MMPs; hold promise as  
488 novel targets for the regulation of macrophage infiltration and subsequent inflammatory responses.  
489 Preliminary work to date has examined elastin and fibrillin-1 fragments which contain repeated  
490 Glycine proline motifs (GxxPG). In mice, neutralization of these GxxPG fragments via antibody  
491 administration reduces macrophage infiltration into the aorta as well as production of MMP2 and  
492 MMP9 (109). However, there has been little if any translational studies concerning the targeting  
493 of ECM fragments for cardiovascular treatment. This may be due to a substantial lack of  
494 knowledge surrounding the interactions between matrix fragments and the adverse inflammatory  
495 response within the myocardium.

496 Over recent years a plethora of matrikines have been recognised for their behaviour-modulating  
497 abilities (summarised in Table 1), yet their precise mechanisms of action in sustaining  
498 inflammation remain to be elucidated. A focus therefore on interactions with ECM and the immune  
499 response, specifically with macrophages within the myocardium is required in future research.  
500 Breakdown of matrix will always be a natural requirement of wound healing and repair, yet  
501 perhaps it is the presence of matrikines or their dysregulation which contributes to adverse

502 remodelling post infarction. Their presence may have to be considered in future targeted therapy  
 503 by means of combined therapy, where both effectors and actuators of remodelling are targeted.  
 504 This is an important consideration going forward, in both therapeutic design, and research models  
 505 of heart failure. Figure 2 depicts the numerous possible therapeutic targets of infarction and  
 506 subsequent heart failure.



507  
 508 **Figure 2 :Potential therapeutic targets of adverse remodelling.** Targets depicted in red boxes and  
 509 include pro-inflammatory cytokines, as well as the CCR2 receptor. Also, to be considered are  
 510 matrikines, which have yet to be assessed in targeted therapy of adverse remodelling.

**Table 1: ECM derived matrikines and their respective modulatory functions**

Identified Cryptid	Function	Source	Reference
GETGPAGPAGPIGPVGARGPA, GPQGPRGDKGETGEQ	Facilitate wound healing enhanced adhesion and antioxidative activities	via cell and	Bovine collagen 1(I) chain (110)
RQVFQVAYIIKA	Facilitate wound healing enhanced cell migration	via cell	$\alpha$ -1 chain (111)

YGDEY	Antioxidant Activity		Tilapia Skin Gelatin hydrolysates	(112)
KNVLVTLYERDEGNLLTEK	Induces production monocytes	MMP9 in	SPARC glycoprotein	(113)
VGVAPG	Induces production fibroblasts	MMP2 in	Elastin	(114)
RGD	Cell adhesion via integrin binding		Fibronectin	(115)
DGGRYY	Activates polymorphonuclear neutrophils		A-1 chain type 1 collagen	(116)
GHK	Chemoattractant for macrophages and mast cells		A-2 chain type 1 collagen	(117)

511

512 **Concluding Remarks and Future Prospects**

513 Undoubtedly, inflammation driven by macrophages plays a key role in heart failure. Numerous  
514 studies have been discussed in this review which pinpoint macrophages as critical mediators of  
515 inflammation and adverse remodelling of ECM. Yet there still remains substantial gaps in our  
516 knowledge of the precise role of macrophages, particularly resident macrophages within the  
517 myocardium. It remains to be established which specific subsets of macrophages are precisely  
518 responsible for the adverse effects of the inflammatory response, and which are necessary for  
519 normal homestatic function. Knockout models which eliminate specific subsets may bring to light  
520 the exact function of resident macrophages, and aid future research in harnessing their protective  
521 nature. As discussed throughout this review, although macrophages are not an active producer of  
522 ECM, they are intimately linked throughout the myocardial milleau in orchestrating remodelling  
523 and deposition; a role that becomes highly prominent following myocardial infarction. Greater  
524 efforts must be made to elucidate the role of the ECM in sustaining activation of macrophages via  
525 matrikines. Further studies of matrix-macrophage communication may reveal not only the precise  
526 mechanisms by which infiltrating macrophages drive remodelling, but also possible novel targets  
527 for future therapies.

528

529

530

531

532

533

534

535

536

537

538 **References**

539

540 1. Benjamin EJ, Blaha MJ, Chiuve SE, Cushman M, Das SR, Deo R, et al. Heart Disease and Stroke  
541 Statistics-2017 Update: A Report From the American Heart Association. *Circulation*. 2017;135(10):e146-  
542 e603.

543 2. Savarese G, Lund LH. Global Public Health Burden of Heart Failure. *Cardiac failure review*.  
544 2017;3(1):7-11.

545 3. Perbellini F, Watson SA, Bardi I, Terracciano CM. Heterocellularity and Cellular Cross-Talk in  
546 the Cardiovascular System. *Frontiers in cardiovascular medicine*. 2018;5:143.

547 4. Wang BX, Kit-Anan W, Terracciano CMN. Many Cells Make Life Work-Multicellularity in  
548 Stem Cell-Based Cardiac Disease Modelling. *International journal of molecular sciences*. 2018;19(11).

549 5. Fountoulaki K, Dargès N, Iliodromitis EK. Cellular Communications in the Heart. *Cardiac failure*  
550 *review*. 2015;1(2):64-8.

551 6. Howard CM, Baudino TA. Dynamic cell-cell and cell-ECM interactions in the heart. *Journal of*  
552 *molecular and cellular cardiology*. 2014;70:19-26.

553 7. Olsson AK, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling - in control of  
554 vascular function. *Nat Rev Mol Cell Biol*. 2006;7(5):359-71.

555 8. Bryant D, Becker L, Richardson J, Shelton J, Franco F, Peshock R, et al. Cardiac failure in  
556 transgenic mice with myocardial expression of tumor necrosis factor-alpha. *Circulation*.  
557 1998;97(14):1375-81.

558 9. Jiang YR, Du JY, Wang DD, Yang X. miRNA-130a improves cardiac function by down-  
559 regulating TNF-alpha expression in a rat model of heart failure. *Eur Rev Med Pharmacol Sci*.  
560 2018;22(23):8454-61.

561 10. Ivey MJ, Tallquist MD. Defining the Cardiac Fibroblast. *Circ J*. 2016;80(11):2269-76.

562 11. Kanekar S, Hirozanne T, Terracio L, Borg TK. Cardiac fibroblasts form and function.  
563 *Cardiovascular pathology : the official journal of the Society for Cardiovascular Pathology*.  
564 1998;7(3):127-33.

565 12. Daskalopoulos EP, Hermans KC, Blankesteyn WM. Cardiac (myo)fibroblast: Novel strategies for  
566 its targeting following myocardial infarction. *Curr Pharm Des*. 2014;20(12):1987-2002.

567 13. Fan D, Takawale A, Lee J, Kassiri Z. Cardiac fibroblasts, fibrosis and extracellular matrix  
568 remodeling in heart disease. *Fibrogenesis & tissue repair*. 2012;5(1):15.

569 14. Murray PJ, Wynn TA. Protective and pathogenic functions of macrophage subsets. *Nature*  
570 *reviews Immunology*. 2011;11(11):723-37.

571 15. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation  
572 and polarization: nomenclature and experimental guidelines. *Immunity*. 2014;41(1):14-20.

573 16. Martinez FO, Sica A, Mantovani A, Locati M. Macrophage activation and polarization. *Frontiers*  
574 *in bioscience : a journal and virtual library*. 2008;13:453-61.

575 17. Yang J, Zhang L, Yu C, Yang X-F, Wang H. Monocyte and macrophage differentiation:  
576 circulation inflammatory monocyte as biomarker for inflammatory diseases. *Biomark Res*. 2014;2(1):1-.

577 18. Mukhopadhyay S, Peiser L, Gordon S. Activation of murine macrophages by *Neisseria*  
578 *meningitidis* and IFN-gamma in vitro: distinct roles of class A scavenger and Toll-like pattern recognition  
579 receptors in selective modulation of surface phenotype. *Journal of leukocyte biology*. 2004;76(3):577-84.

580 19. Ferrante CJ, Pinhal-Enfield G, Elson G, Cronstein BN, Hasko G, Outram S, et al. The adenosine-  
581 dependent angiogenic switch of macrophages to an M2-like phenotype is independent of interleukin-4  
582 receptor alpha (IL-4Ralpha) signaling. *Inflammation*. 2013;36(4):921-31.

583 20. Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that  
584 have ingested apoptotic cells in vitro inhibit proinflammatory cytokine production through  
585 autocrine/paracrine mechanisms involving TGF-beta, PGE2, and PAF. *J Clin Invest*. 1998;101(4):890-8.

586 21. Martin CJ, Peters KN, Behar SM. Macrophages clean up: efferocytosis and microbial control.  
587 *Curr Opin Microbiol*. 2014;17:17-23.



- 588 22. Davies LC, Jenkins SJ, Allen JE, Taylor PR. Tissue-resident macrophages. *Nature immunology*.  
589 2013;14(10):986-95.
- 590 23. Chen B, Brickshawana A, Frangogiannis NG. The Functional Heterogeneity of Resident Cardiac  
591 Macrophages in Myocardial Injury CCR2(+) Cells Promote Inflammation, Whereas CCR2(-) Cells  
592 Protect. *Circulation research*. 2019;124(2):183-5.
- 593 24. Honold L, Nahrendorf M. Resident and Monocyte-Derived Macrophages in Cardiovascular  
594 Disease. 2018;122(1):113-27.
- 595 25. Bajpai G, Schneider C, Wong N, Bredemeyer A, Hulsmans M, Nahrendorf M, et al. The human  
596 heart contains distinct macrophage subsets with divergent origins and functions. *Nature Medicine*.  
597 2018;24(8):1234-45.
- 598 26. Epelman S, Lavine Kory J, Randolph Gwendalyn J. Origin and Functions of Tissue  
599 Macrophages. *Immunity*. 2014;41(1):21-35.
- 600 27. Leid J, Carrelha J, Boukarabila H, Epelman S, Jacobsen SE, Lavine KJ. Primitive Embryonic  
601 Macrophages are Required for Coronary Development and Maturation. *Circulation research*.  
602 2016;118(10):1498-511.
- 603 28. Bajpai G, Bredemeyer A, Li W, Zaitsev K, Koenig AL, Lokshina I, et al. Tissue Resident CCR2-  
604 and CCR2+ Cardiac Macrophages Differentially Orchestrate Monocyte Recruitment and Fate  
605 Specification Following Myocardial Injury. *Circulation research*. 2019;124(2):263-78.
- 606 29. Dick SA, Macklin JA, Nejat S, Momen A, Clemente-Casares X, Althagafi MG, et al. Self-  
607 renewing resident cardiac macrophages limit adverse remodeling following myocardial infarction. *Nature*  
608 *immunology*. 2019;20(1):29-39.
- 609 30. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages Facilitate  
610 Electrical Conduction in the Heart. *Cell*. 2017;169(3):510-22.e20.
- 611 31. Panek CA, Ramos MV, Mejias MP, Abrey-Recalde MJ, Fernandez-Brando RJ, Gori MS, et al.  
612 Differential expression of the fractalkine chemokine receptor (CX3CR1) in human monocytes during  
613 differentiation. *Cellular & molecular immunology*. 2015;12(6):669-80.
- 614 32. Nielsen SH, Mouton AJ, DeLeon-Pennell KY, Genovese F, Karsdal M, Lindsey ML.  
615 Understanding cardiac extracellular matrix remodeling to develop biomarkers of myocardial infarction  
616 outcomes. *Matrix Biology*. 2019;75-76:43-57.
- 617 33. Frangogiannis NG. The extracellular matrix in myocardial injury, repair, and remodeling. *J Clin*  
618 *Invest*. 2017;127(5):1600-12.
- 619 34. Mizuno T, Mickle DA, Kiani CG, Li RK. Overexpression of elastin fragments in infarcted  
620 myocardium attenuates scar expansion and heart dysfunction. *American journal of physiology Heart and*  
621 *circulatory physiology*. 2005;288(6):H2819-27.
- 622 35. Lockhart M, Wirrig E, Phelps A, Wessels A. Extracellular matrix and heart development. *Birth*  
623 *Defects Res A Clin Mol Teratol*. 2011;91(6):535-50.
- 624 36. Chen WY, Hong J, Gannon J, Kakkar R, Lee RT. Myocardial pressure overload induces systemic  
625 inflammation through endothelial cell IL-33. *Proceedings of the National Academy of Sciences of the*  
626 *United States of America*. 2015;112(23):7249-54.
- 627 37. Entman ML, Michael L, Rossen RD, Dreyer WJ, Anderson DC, Taylor AA, et al. Inflammation  
628 in the course of early myocardial ischemia. *FASEB journal : official publication of the Federation of*  
629 *American Societies for Experimental Biology*. 1991;5(11):2529-37.
- 630 38. Ferrucci L, Fabbri E. Inflammageing: chronic inflammation in ageing, cardiovascular disease,  
631 and frailty. *Nature reviews Cardiology*. 2018;15(9):505-22.
- 632 39. Pagano F, Angelini F, Castaldo C, Picchio V, Messina E, Sciarretta S, et al. Normal versus  
633 Pathological Cardiac Fibroblast-Derived Extracellular Matrix Differentially Modulates Cardiosphere-  
634 Derived Cell Paracrine Properties and Commitment. *Stem cells international*. 2017;2017:7396462.
- 635 40. van Nieuwenhoven FA, Munts C, Op't Veld RC, Gonzalez A, Diez J, Heymans S, et al. Cartilage  
636 intermediate layer protein 1 (CILP1): A novel mediator of cardiac extracellular matrix remodelling.  
637 *Scientific reports*. 2017;7(1):16042.

638 41. Anversa P, Sonnenblick EH. Ischemic cardiomyopathy: Pathophysiologic mechanisms. Progress  
639 in Cardiovascular Diseases. 1990;33(1):49-70.

640 42. Isomi M, Sadahiro T, Ieda M. Progress and Challenge of Cardiac Regeneration to Treat Heart  
641 Failure. Journal of cardiology. 2019;73(2):97-101.

642 43. Frangogiannis NG. Inflammation in cardiac injury, repair and regeneration. Current opinion in  
643 cardiology. 2015;30(3):240-5.

644 44. Cohn JN, Ferrari R, Sharpe N. Cardiac remodeling--concepts and clinical implications: a  
645 consensus paper from an international forum on cardiac remodeling. Behalf of an International Forum on  
646 Cardiac Remodeling. Journal of the American College of Cardiology. 2000;35(3):569-82.

647 45. Heidt T, Courties G, Dutta P, Sager HB, Sebas M, Iwamoto Y, et al. Differential contribution of  
648 monocytes to heart macrophages in steady-state and after myocardial infarction. Circulation research.  
649 2014;115(2):284-95.

650 46. Ma Y, Mouton AJ, Lindsey ML. Cardiac macrophage biology in the steady-state heart, the aging  
651 heart, and following myocardial infarction. Translational research : the journal of laboratory and clinical  
652 medicine. 2018;191:15-28.

653 47. Nahrendorf M, Swirski FK, Aikawa E, Stangenberg L, Wurdinger T, Figueiredo JL, et al. The  
654 healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary  
655 functions. The Journal of experimental medicine. 2007;204(12):3037-47.

656 48. Mouton AJ, DeLeon-Pennell KY, Rivera Gonzalez OJ, Flynn ER, Freeman TC, Saucerman JJ, et  
657 al. Mapping macrophage polarization over the myocardial infarction time continuum. Basic research in  
658 cardiology. 2018;113(4):26.

659 49. Galvan-Pena S, O'Neill LA. Metabolic reprogramming in macrophage polarization. Frontiers in  
660 immunology. 2014;5:420.

661 50. Poon IKH, Lucas CD, Rossi AG, Ravichandran KS. Apoptotic cell clearance: basic biology and  
662 therapeutic potential. Nature Reviews Immunology. 2014;14:166.

663 51. Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that  
664 have ingested apoptotic cells in vitro inhibit proinflammatory cytokine production through  
665 autocrine/paracrine mechanisms involving TGF-beta, PGE2, and PAF. The Journal of Clinical  
666 Investigation. 1998;101(4):890-8.

667 52. Zhang S, Weinberg S, DeBerge M, Gainullina A, Schipma M, Kinchen JM, et al. Efferocytosis  
668 Fuels Requirements of Fatty Acid Oxidation and the Electron Transport Chain to Polarize Macrophages  
669 for Tissue Repair. Cell Metabolism. 2019;29(2):443-56.e5.

670 53. Yurdagul A, Jr., Doran AC, Cai B, Fredman G, Tabas IA. Mechanisms and Consequences of  
671 Defective Efferocytosis in Atherosclerosis. Frontiers in cardiovascular medicine. 2017;4:86.

672 54. Gruzdeva O, Uchasova E, Dyleva Y, Akbasheva O, Matveeva V, Karetnikova V, et al.  
673 Relationship key factor of inflammation and the development of complications in the late period of  
674 myocardial infarction in patients with visceral obesity. BMC cardiovascular disorders. 2017;17(1):36-  
675 55. Tilg H, Moschen AR. Inflammatory mechanisms in the regulation of insulin resistance.  
676 Molecular medicine (Cambridge, Mass). 2008;14(3-4):222-31.

677 56. Lu P, Takai K, Weaver VM, Werb Z. Extracellular matrix degradation and remodeling in  
678 development and disease. Cold Spring Harbor perspectives in biology. 2011;3(12).

679 57. Aimes RT, Quigley JP. Matrix metalloproteinase-2 is an interstitial collagenase. Inhibitor-free  
680 enzyme catalyzes the cleavage of collagen fibrils and soluble native type I collagen generating the  
681 specific 3/4- and 1/4-length fragments. The Journal of biological chemistry. 1995;270(11):5872-6.

682 58. Barker TH, Engler AJ. The provisional matrix: setting the stage for tissue repair outcomes.  
683 Matrix biology : journal of the International Society for Matrix Biology. 2017;60-61:1-4.

684 59. Flick MJ, Du X, Witte DP, Jirouskova M, Soloviev DA, Busuttill SJ, et al. Leukocyte engagement  
685 of fibrin(ogen) via the integrin receptor alphaMbeta2/Mac-1 is critical for host inflammatory response in  
686 vivo. J Clin Invest. 2004;113(11):1596-606.

687 60. Martino MM, Briquez PS, Ranga A, Lutolf MP, Hubbell JA. Heparin-binding domain of  
688 fibrin(ogen) binds growth factors and promotes tissue repair when incorporated within a synthetic matrix.  
689 Proceedings of the National Academy of Sciences of the United States of America. 2013;110(12):4563-8.  
690 61. Creemers E, Cleutjens J, Smits J, Heymans S, Moons L, Collen D, et al. Disruption of the  
691 Plasminogen Gene in Mice Abolishes Wound Healing after Myocardial Infarction. The American Journal  
692 of Pathology. 2000;156(6):1865-73.  
693 62. Motley MP, Madsen DH, Jurgensen HJ, Spencer DE, Szabo R, Holmbeck K, et al. A CCR2  
694 macrophage endocytic pathway mediates extravascular fibrin clearance in vivo. Blood.  
695 2016;127(9):1085-96.  
696 63. Hsieh JY, Smith TD, Meli VS, Tran TN, Botvinick EL, Liu WF. Differential regulation of  
697 macrophage inflammatory activation by fibrin and fibrinogen. Acta Biomaterialia. 2017;47:14-24.  
698 64. Rucklidge GJ, Milne G, McGaw BA, Milne E, Robins SP. Turnover rates of different collagen  
699 types measured by isotope ratio mass spectrometry. Biochimica et biophysica acta. 1992;1156(1):57-61.  
700 65. Gonzalez-Santamaria J, Villalba M, Busnadiego O, Lopez-Olaneta MM, Sandoval P, Snabel J, et  
701 al. Matrix cross-linking lysyl oxidases are induced in response to myocardial infarction and promote  
702 cardiac dysfunction. Cardiovascular research. 2016;109(1):67-78.  
703 66. Blackburn NJR, Vulesevic B, McNeill B, Cimenci CE, Ahmadi A, Gonzalez-Gomez M, et al.  
704 Methylglyoxal-derived advanced glycation end products contribute to negative cardiac remodeling and  
705 dysfunction post-myocardial infarction. Basic research in cardiology. 2017;112(5):57.  
706 67. Richardson WJ, Clarke SA, Quinn TA, Holmes JW. Physiological Implications of Myocardial  
707 Scar Structure. Compr Physiol. 2015;5(4):1877-909.  
708 68. Kaikita K, Hayasaki T, Okuma T, Kuziel WA, Ogawa H, Takeya M. Targeted deletion of CC  
709 chemokine receptor 2 attenuates left ventricular remodeling after experimental myocardial infarction. Am  
710 J Pathol. 2004;165(2):439-47.  
711 69. Huleihel M, Douvdevani A, Segal S, Apte RN. Regulation of interleukin 1 generation in immune-  
712 activated fibroblasts. European journal of immunology. 1990;20(4):731-8.  
713 70. Halade GV, Jin YF, Lindsey ML. Matrix metalloproteinase (MMP)-9: a proximal biomarker for  
714 cardiac remodeling and a distal biomarker for inflammation. Pharmacology & therapeutics.  
715 2013;139(1):32-40.  
716 71. Zavadzka JA, Mukherjee R, Rivers WT, Patel RK, Meyer EC, Black LE, et al. Direct regulation  
717 of membrane type 1 matrix metalloproteinase following myocardial infarction causes changes in survival,  
718 cardiac function, and remodeling. American journal of physiology Heart and circulatory physiology.  
719 2011;301(4):H1656-66.  
720 72. Leask A. Potential Therapeutic Targets for Cardiac Fibrosis. 2010;106(11):1675-80.  
721 73. Dobaczewski M, Bujak M, Li N, Gonzalez-Quesada C, Mendoza LH, Wang XF, et al. Smad3  
722 signaling critically regulates fibroblast phenotype and function in healing myocardial infarction.  
723 Circulation research. 2010;107(3):418-28.  
724 74. Saxena A, Chen W, Su Y, Rai V, Uche OU, Li N, et al. IL-1 induces proinflammatory leukocyte  
725 infiltration and regulates fibroblast phenotype in the infarcted myocardium. Journal of immunology  
726 (Baltimore, Md : 1950). 2013;191(9):4838-48.  
727 75. van Nieuwenhoven FA, Hemmings KE, Porter KE, Turner NA. Combined effects of interleukin-  
728 alpha and transforming growth factor-beta1 on modulation of human cardiac fibroblast function. Matrix  
729 Biol. 2013;32(7-8):399-406.  
730 76. Walker GA, Masters KS, Shah DN, Anseth KS, Leinwand LA. Valvular myofibroblast activation  
731 by transforming growth factor-beta: implications for pathological extracellular matrix remodeling in heart  
732 valve disease. Circulation research. 2004;95(3):253-60.  
733 77. Sahin H, Wasmuth HE. Chemokines in tissue fibrosis. Biochimica et Biophysica Acta (BBA) -  
734 Molecular Basis of Disease. 2013;1832(7):1041-8.  
735 78. Thompson RW, Pesce JT, Ramalingam T, Wilson MS, White S, Cheever AW, et al. Cationic  
736 Amino Acid Transporter-2 Regulates Immunity by Modulating Arginase Activity. PLOS Pathogens.  
2008;4(3):e1000023.

738 79. Fontes JA, Rose NR, Čiháková D. The varying faces of IL-6: From cardiac protection to cardiac  
739 failure. *Cytokine*. 2015;74(1):62-8.

740 80. Mayfield AE, Kanda P, Nantsios A, Parent S, Mount S, Dixit S, et al. Interleukin-6 Mediates  
741 Post-Infarct Repair by Cardiac Explant-Derived Stem Cells. *Theranostics*. 2017;7(19):4850-61.

742 81. Müller J, Gorresen S, Grandoch M, Feldmann K, Kretschmer I, Lehr S, et al. Interleukin-6-  
743 dependent phenotypic modulation of cardiac fibroblasts after acute myocardial infarction. *Basic research*  
744 *in cardiology*. 2014;109(6):440.

745 82. Verma Suresh K, Krishnamurthy P, Barefield D, Singh N, Gupta R, Lambers E, et al. Interleukin-  
746 10 Treatment Attenuates Pressure Overload-Induced Hypertrophic Remodeling and Improves Heart  
747 Function via Signal Transducers and Activators of Transcription 3-Dependent Inhibition of Nuclear  
748 Factor- $\kappa$ B. *Circulation*. 2012;126(4):418-29.

749 83. Giugliano GR, Giugliano RP, Gibson CM, Kuntz RE. Meta-analysis of corticosteroid treatment in  
750 acute myocardial infarction. *The American journal of cardiology*. 2003;91(9):1055-9.

751 84. Gong K, Zhang Z, Sun X, Zhang X, Li A, Yan J, et al. The nonspecific anti-inflammatory therapy  
752 with methotrexate for patients with chronic heart failure. *American heart journal*. 2006;151(1):62-8.

753 85. Moreira DM, Lueneberg ME, da Silva RL, Fattah T, Gottschall CAM. Methotrexate Therapy in  
754 ST-Segment Elevation Myocardial Infarction: A Randomized Double-Blind, Placebo-Controlled Trial  
755 (TETHYS Trial). *Journal of cardiovascular pharmacology and therapeutics*. 2017;22(6):538-45.

756 86. Moreira DM, Vieira JL, Gottschall CA. The effects of Methotrexate therapy on the physical  
757 capacity of patients with Ischemic heart failure: a randomized double-blind, placebo-controlled trial  
758 (METIS trial). *Journal of cardiac failure*. 2009;15(10):828-34.

759 87. Hausenloy DJ, Boston-Griffiths EA, Yellon DM. Cyclosporin A and cardioprotection: from  
760 investigative tool to therapeutic agent. *British journal of pharmacology*. 2012;165(5):1235-45.

761 88. Yingzhong C, Lin C, Chunbin W. Clinical effects of cyclosporine A on reperfusion injury in  
762 myocardial infarction: a meta-analysis of randomized controlled trials. *SpringerPlus*. 2016;5(1):1117.

763 89. Sager HB, Heidt T, Hulsmans M, Dutta P, Courties G, Sebas M, et al. Targeting Interleukin-1beta  
764 Reduces Leukocyte Production After Acute Myocardial Infarction. *Circulation*. 2015;132(20):1880-90.

765 90. Toldo S, Mezzaroma E, Van Tassell BW, Farkas D, Marchetti C, Voelkel NF, et al. Interleukin-  
766 1beta blockade improves cardiac remodelling after myocardial infarction without interrupting the  
767 inflammasome in the mouse. *Experimental physiology*. 2013;98(3):734-45.

768 91. Abbate A, Kontos MC, Grizzard JD, Biondi-Zoccai GG, Van Tassell BW, Robati R, et al.  
769 Interleukin-1 blockade with anakinra to prevent adverse cardiac remodeling after acute myocardial  
770 infarction (Virginia Commonwealth University Anakinra Remodeling Trial [VCU-ART] Pilot study). *The*  
771 *American journal of cardiology*. 2010;105(10):1371-7.e1.

772 92. Morton AC, Rothman AM, Greenwood JP, Gunn J, Chase A, Clarke B, et al. The effect of  
773 interleukin-1 receptor antagonist therapy on markers of inflammation in non-ST elevation acute coronary  
774 syndromes: the MRC-ILA Heart Study. *European heart journal*. 2015;36(6):377-84.

775 93. Abbate A, Van Tassell BW, Biondi-Zoccai G, Kontos MC, Grizzard JD, Spillman DW, et al.  
776 Effects of interleukin-1 blockade with anakinra on adverse cardiac remodeling and heart failure after  
777 acute myocardial infarction [from the Virginia Commonwealth University-Anakinra Remodeling Trial (2)  
778 (VCU-ART2) pilot study]. *The American journal of cardiology*. 2013;111(10):1394-400.

779 94. Ridker PM, MacFadyen JG, Everett BM, Libby P, Thuren T, Glynn RJ. Relationship of C-  
780 reactive protein reduction to cardiovascular event reduction following treatment with canakinumab: a  
781 secondary analysis from the CANTOS randomised controlled trial. *Lancet (London, England)*.  
782 2018;391(10118):319-28.

783 95. Padfield GJ, Din JN, Koushiappi E, Mills NL, Robinson SD, Cruden Nle M, et al. Cardiovascular  
784 effects of tumour necrosis factor alpha antagonism in patients with acute myocardial infarction: a first in  
785 human study. *Heart (British Cardiac Society)*. 2013;99(18):1330-5.

786 96. Mann DL, McMurray JJ, Packer M, Swedberg K, Borer JS, Colucci WS, et al. Targeted  
787 anticytokine therapy in patients with chronic heart failure: results of the Randomized Etanercept  
788 Worldwide Evaluation (RENEWAL). *Circulation*. 2004;109(13):1594-602.

789 97. Struthers M, Pasternak A. CCR2 antagonists. *Current topics in medicinal chemistry*.  
790 2010;10(13):1278-98.

791 98. Sager HB, Dutta P, Dahlman JE, Hulsmans M, Courties G, Sun Y, et al. RNAi targeting multiple  
792 cell adhesion molecules reduces immune cell recruitment and vascular inflammation after myocardial  
793 infarction. *Science translational medicine*. 2016;8(342):342ra80.

794 99. Leuschner F, Dutta P, Gorbato R, Novobrantseva TI, Donahoe JS, Courties G, et al. Therapeutic  
795 siRNA silencing in inflammatory monocytes in mice. *Nature Biotechnology*. 2011;29:1005.

796 100. Majmudar MD, Keliher EJ, Heidt T, Leuschner F, Truelove J, Sena BF, et al. Monocyte-directed  
797 RNAi targeting CCR2 improves infarct healing in atherosclerosis-prone mice. *Circulation*.  
798 2013;127(20):2038-46.

799 101. Monaghan M, Browne S, Schenke-Layland K, Pandit A. A Collagen-based Scaffold Delivering  
800 Exogenous MicroRNA-29B to Modulate Extracellular Matrix Remodeling. *Molecular Therapy*.  
801 2014;22(4):786-96.

802 102. Monaghan MG, Holeiter M, Brauchle E, Layland SL, Lu Y, Deb A, et al. Exogenous miR-29B  
803 Delivery Through a Hyaluronan-Based Injectable System Yields Functional Maintenance of the Infarcted  
804 Myocardium. *Tissue Engineering Part A*. 2017;24(1-2):57-67.

805 103. Panahi M, Papanikolaou A, Torabi A, Zhang J-G, Khan H, Vazir A, et al. Immunomodulatory  
806 interventions in myocardial infarction and heart failure: a systematic review of clinical trials and meta-  
807 analysis of IL-1 inhibition. *Cardiovascular research*. 2018;114(11):1445-61.

808 104. Parks WC, Wilson CL, Lopez-Boado YS. Matrix metalloproteinases as modulators of  
809 inflammation and innate immunity. *Nature reviews Immunology*. 2004;4(8):617-29.

810 105. Maquart FX, Bellon G, Pasco S, Monboisse JC. Matrikines in the regulation of extracellular  
811 matrix degradation. *Biochimie*. 2005;87(3-4):353-60.

812 106. Adair-Kirk TL, Senior RM. Fragments of extracellular matrix as mediators of inflammation. *The*  
813 *international journal of biochemistry & cell biology*. 2008;40(6-7):1101-10.

814 107. Kwon YW, Heo SC, Lee TW, Park GT, Yoon JW, Jang IH, et al. N-Acetylated Proline-Glycine-  
815 Proline Accelerates Cutaneous Wound Healing and Neovascularization by Human Endothelial Progenitor  
816 Cells. *Scientific reports*. 2017;7:43057.

817 108. Weathington NM, van Houwelingen AH, Noerager BD, Jackson PL, Kraneveld AD, Galin FS, et  
818 al. A novel peptide CXCR ligand derived from extracellular matrix degradation during airway  
819 inflammation. *Nat Med*. 2006;12(3):317-23.

820 109. Guo G, Munoz-Garcia B, Ott CE, Grunhagen J, Mousa SA, Pletschacher A, et al. Antagonism of  
821 GxxPG fragments ameliorates manifestations of aortic disease in Marfan syndrome mice. *Human*  
822 *molecular genetics*. 2013;22(3):433-43.

823 110. Banerjee P, Suseela G, Shanthi C. Isolation and identification of cryptic bioactive regions in  
824 bovine achilles tendon collagen. *The protein journal*. 2012;31(5):374-86.

825 111. Malinda KM, Wysocki AB, Koblinski JE, Kleinman HK, Ponce ML. Angiogenic laminin-derived  
826 peptides stimulate wound healing. *The international journal of biochemistry & cell biology*.  
827 2008;40(12):2771-80.

828 112. Zhang Y, Duan X, Zhuang Y. Purification and characterization of novel antioxidant peptides  
829 from enzymatic hydrolysates of tilapia (*Oreochromis niloticus*) skin gelatin. *Peptides*. 2012;38(1):13-21.

830 113. Shankavaram UT, DeWitt DL, Funk SE, Sage EH, Wahl LM. Regulation of human monocyte  
831 matrix metalloproteinases by SPARC. *Journal of cellular physiology*. 1997;173(3):327-34.

832 114. Brassart B, Randoux A, Hornebeck W, Emonard H. Regulation of matrix metalloproteinase-2  
833 (gelatinase A, MMP-2), membrane-type matrix metalloproteinase-1 (MT1-MMP) and tissue inhibitor of  
834 metalloproteinases-2 (TIMP-2) expression by elastin-derived peptides in human HT-1080 fibrosarcoma  
835 cell line. *Clinical & experimental metastasis*. 1998;16(6):489-500.

836 115. Mooradian DL, McCarthy JB, Skubitz AP, Cameron JD, Furcht LT. Characterization of FN-C/H-  
837 V, a novel synthetic peptide from fibronectin that promotes rabbit corneal epithelial cell adhesion,  
838 spreading, and motility. *Investigative ophthalmology & visual science*. 1993;34(1):153-64.

839 116. Monboisse JC, Bellon G, Randoux A, Dufer J, Borel JP. Activation of human neutrophils by type  
840 I collagen. Requirement of two different sequences. The Biochemical journal. 1990;270(2):459-62.  
841 117. Zetter BR, Rasmussen N, Brown L. An in vivo assay for chemoattractant activity. Laboratory  
842 investigation; a journal of technical methods and pathology. 1985;53(3):362-8.

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

863

864

865

866